1. **The impact of vaping on adolescent lung function and nasal epithelium gene expression.**
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**Abstract**

**Background:** Electronic cigarettes (e-cigs) are touted as safer alternatives to traditional tobacco products; however, there are no substantive data to corroborate this claim. Vaping among adolescents is a pressing public health problem and is a risk factor for negative health effects. We aimed to investigate the impact of vape exposure on lung function and nasal epithelial gene expression in adolescents. We hypothesized that vape exposure results in abnormal lung function and differential expression of inflammatory genes in the nasal epithelium of vape users in adolescence.

**Study Design and Methods:** We performed focus groups and interviews of 50 middle and high school students at high school and community-based settings in Colorado to explore youth perceptions on vaping. Participants completed focus groups and confidential surveys on their vaping use. Airflow obstruction was investigated using impulse oscillometry (IOS) and spirometry. Nasal epithelial brushing was collected for gene expression analyses. RNA-sequencing was performed using the Illumina HiSeq platform. We also performed a targeted DNA methylation analysis using Illumina’s methylation platform of the differentially expressed genes from the RNA-seq data. We compared IOS measures between vaping and non-vaping groups using T-tests. Differential expression testing was performed in DESeq2 and enrichment was performed using Gene Set Enrichment Analysis (GSEA). Written informed consent was obtained from participants and Institutional Review Board approval was obtained.

**Results:** Fifty-one participants were recruited. Vaping subjects were defined as those who reported vaping within the past 6 months (hereafter vape users) while non-vaping control subjects did not report any vape exposure in the past 6 months (hereafter non-vape users). One subject did not report vape status. The mean (standard deviation) age was 15.1 (1.5) years for vape users and 14.5 (1.4) years for non-vape users. We found significant differences in airway resistance by vaping status. Mean airway resistance identified by the IOS R5 measure revealed that vape users (n=13) had higher values than non-vape users (n=37) [p=0.026], an early indication of increased airflow obstruction. Of 16,860 nasal epithelial genes tested, 7,136 were significantly differentially expressed between vape and non-vape users (false discovery rate FDR < 0.05), after covariate adjustment. Enrichment analyses identified overexpression of inflammatory response genes and underexpression of genes associated with ciliogenesis in the vape users compared to non-vape users. DNA methylation analysis revealed that REXO1 (FDR=0.01) and CERK (FDR=0.06) were differentially methylated among vape users compared to non-vape users.

**Interpretation:** Our results suggest that vaping increases airway resistance and dysresgulation of nasal epithelial genes, including genes involved in airway inflammation and ciliary function. Furthermore, dysregulation of these gene expression pathways may be a biological mechanism for the development of increased airway resistance due to vaping even during adolescence.

**Clinical Trial Registration:** N/A

1. **Background**

The increasing use of Electronic Nicotine Delivery Systems (ENDS), such as electronic cigarettes (e-cigs), is a significant and emerging public health problem [1]. ENDS represents a diverse class of products such as electronic cigarettes, vapes, vaporizers, vape pens, hookah pens, and pods [2], and exposure to ENDS aerosols depends on the user and device characteristics [2]. They may contain nicotine and can be classified as tobacco products [3]. Herein, we focus on one type of ENDS, namely e-cigs, the use of which we term vaping. E-cigs are touted as safe alternatives to traditional tobacco products, however, there is no substantive data to corroborate this assertion [4]. The solutions in e-cigs, and their resulting aerosols, contain nicotine, carcinogens, and metal particles, to which users and nonusers in close proximity can be exposed [3]. The youth and young adults are the predominant population using e-cigarettes and are at the greatest risk to the negative effects of nicotine exposure [5]. Given the attractive flavors, marketing and design, and its appeal to youth, e-cigs have the potential to reverse decades of progress achieved in nicotine and tobacco product use [6].

While traditional cigarette smoking has declined, e-cig usage (or vape use) has increased, attracting both former, current and never tobacco smokers [1]. Vaping among youth is a pressing public health issue, with prevalence of use surpassing that of tobacco cigarettes [7]. There is a reported increase in past-30-day e-cigarette use among high school students. Particularly, the National Youth Tobacco Survey reported increased from 11.3% in 2017 to 20.8% in 2018 [8]. Monitoring the Future Study also reported increased use from 11.0% in 2017 to 20.9% in 2018 [9]. A recent study on failed nicotine quit attempts, among US adolescent e-cig and traditional cigarette users, shows that levels are back to where it was 13 years ago [10]. Certainly, the contribution of e-cigs to unsuccessful nicotine quit attempts among adolescents is substantial and warrants urgent attention.

The pathophysiologic impacts of e-cig exposure on the human pulmonary system are being elucidated.Tobacco smoke is known to be highly proinflammatory and has been shown to trigger the release of inflammatory cytokines and other biological changes, including goblet cell metaplasia and neutrophil influx [11], however the impact of e-cig aerosols in the long term is not clear. The solutions found in e-cigarettes present a novel mixture of chemicals, including flavors and sweeteners designed to mask nicotine’s bitter taste [1]. Thus, while nicotine is known to adversely alter airway physiology, the effects of these novel chemical mixtures, either by itself, and/or together with nicotine have not been studied. Short-term exposure to e-cigarettes in healthy adults increases airway resistance, with some evidence demonstrating decreased lung function after exposure [12]. However, what happens in the long term during a critical development window such as adolescence, is unknown.

Given the paucity of information on the effects of vaping on the lungs in the long-term, the high-risk youth population with access to these devices, and the current epidemic, we recruited adolescents who vape to help address these knowledge gaps. By presenting evidence on biologic determinants of reduced lung function we will fill in a critical knowledge gap on the health effects of vaping in the human population. Thus, we aimed to investigate the impact of vape exposure on measures of lung function in adolescents and to compare nasal epithelial gene expression and DNA methylation in those who vape to non-vaping controls to determine the biological impact of vape exposure. The motivating hypothesis for this work is that vape exposure is associated with abnormal lung function measures and that nasal epithelial gene expression would be modified by vape exposure. To address this hypothesis, we sought to determine the impact of vape exposure on measures of lung function and its impact on nasal epithelial gene expression in adolescents who vape.

**Methods**

**Study Participants**

Study participants were enrolled in a pilot study aimed to examine vaping initiation and its respiratory effects among youth in Colorado. Adolescent participants completed focus groups on reasons for vaping initiation and confidential surveys on their vaping use. Adolescents from high schools in Pueblo, Aurora and Denver were enrolled in this pilot study. Specifically, youth aged 12 to 17 years were eligible as participants. Written informed consent was obtained from each of the participants. We used Research Electronic Data Capture (REDCap) to securely enter and store data. The Institutional Review Board at the University of Colorado Anschutz Medical Campus approved the current study.

**Vape use (or vaping) variable definition.**

Subjects who reported vape use in the last six months were considered to be vapers (hereafter vape users), while those who did not report vape use in the last six months were considered as control subjects (hereafter non-vape users).

**Impulse Oscillometry**

Impulse oscillometry (IOS) measurements were obtained from each participant. Since IOS permits passive measurement of lung mechanics, it can identify small airway obstruction and is more sensitive than spirometry for peripheral airway disease [13, 14]. Using tremoflo, a portable IOS analyzer (Thorasys INC, Montreal, Canada), forced oscillation measurements (FOT) measurements were applied in 60 second measurements in triplicate, capturing reactance (XR), resistance (R), resonance frequency (XA). Data were captured within 5–37 Hz [15]. Additionally, we performed spirometry at each visit following the American Thoracic Society/European Respiratory Society guidelines [16, 17].

**Nasal epithelium gene expression**

RNA was isolated from nasal epithelial brush specimens obtained from our study participants.Vap at the Genomics Core at the University of Colorado. To minimize potential batch effects, all samples were submitted together for RNA-seq. We examined the quality of the sequencing reads (FastQC[18]) and removed adaptors and low quality base calls (Cutadapt [19]). Sequences were aligned to the human genome (GENCODE GRCh38) [20] and read counts overlapping each gene were reported using STAR [21]. We performed quality control at the sample and gene-level, removing genes detected with 0 reads in ≥ 75% of samples or with a range of reads ≤ 100.

**DNA methylation**

DNA was isolated and purified from participants nasal samples using the Maxwell® 16 Integrated System (Promega Corp, Madison, WI) [22]. We assessed genome-wide DNA methylation using the Illumina Infinium Human Methylation 850K beadchip profiling microarray (Illumina Inc, San Diego, CA). Samples were analyzed at the Genomics Core at the University of Colorado and were processed under standardized conditions and processed in the same batch. Illumina results were represented as average β values (methylated probe intensity over sum of methylated and unmethylated probe intensities). M-values were calculated as the log2 ratio of the intensities of methylated probe versus unmethylated probe and used in subsequent analysis.

**Covariates**

### For adjusted analyses, we include the following the covariates into the models, as these have been identified as potential confounders in the literature: 1) Recruiting center (Pueblo, Denver/Commerce City, Aurora), which encompasses a broader geographic region where the participant lives, 2) Age, 3) Sex. Due to some confusion between the sex and gender fields in study questionnaire, we used methylation data to identify and confirm the biological sex of all individuals.

**Statistical Analysis**

All analyses were performed using R version 4.2.1. We examined differences in demographic measures among vape and non-vape users using Fisher’s Exacttests for categorical variables and two-sample t-tests for continuous variables. Next, we conducted a series of bioinformatic analyses to evaluate associations between vape status and lung function and gene expression among study participants. For the lung function data, IOS measures were visually inspected for normality using histograms before conducting association studies with vaping status.

Gene expression data normalization  
To correct for unwanted technical effects, we used the between-sample normalization of Removal of Unwanted Variance using residuals (RUVr) from a first pass generalized linear model (GLM) including vape status, sex, and age [23]. Two normalization factors were included for RUVr after visual inspection of an elbow plot, relative log expression plots, and dendrograms. This analysis used edgeR to fit the first pass GLM [24].

Gene counts were then modeled using a series of negative binomial models from the R package *DESeq2* to preform Likelihood Ratio Tests (LRT) [25]. All models adjusted for sex, age, recruitment center, and the normalization factors. The main test of interest for each gene evaluated whether including vape status significantly increased model fit while also adjusting for recruitment center, age, and sex. The other resulting LRTs investigate the presence of recruitment center bias in the data. To correct for multiple testing the Benjamini and Hochberg False Discovery Rate (FDR) adjustment was used. Significance was then set at FDR < 0.05. In addition, we focused on genes with an fold change (FC) effect cutoff of |Log2(FC)| > 2.

**Sensitivity analysis**

To check the robustness of our results, we conducted a sensitivity analysis of only subjects from the Pueblo recruitment center (where 12/13 vape users were recruited). An LRT was fit to determine if *vape status* contributes significantly to gene expression differences with only the subjects from the Pueblo recruitment center while adjusting for age, sex, and the 2 normalization factors. FDR was used to correct for multiple testing and significance was set at FDR ≤ 0.05 and a magnitude of effect cutoff of |Log2(FC)| > 2.

**Enrichment analysis**

To identify differentially enriched biological pathways or ontologies, Gene Set Enrichment Analysis (GSEA) was conducted after obtaining differential gene expression results using the R package *fGSEA* ver. 1.23.0.[26] To compare across the pathway or ontology, data base genes were mapped to Entrez (NCBI) IDs. Gene ranks were based on the direction of the FC times -log10 of the nominal p-value from the LRT [27].

**Databases**

We implemented GSEA for Reactome, Gene Ontology (GO), and Koyoto Encyclopedia of Genes and Genomes (KEGG). Only pathways or ontologies with 2 or more unique Entrez genes were considered for inclusion in GSEA. After the initial analysis, we filtered redundant pathways using the function *collapsePathways* from fGSEA.[28] Pathways or ontologies that are remaining after filtering other pathways or ontologies are referred to as non-redundant. All pathways were retrieved from Ensembl (ver. 106) using biomaRt ver. 2.53.2.[29, 30].

**DNA methylation analysis**

We mapped CpG sites to their respective genes using annotation for Illumina’s 850K methylation arrays [31] to generate a list of CpGs for targeted methylation analysis. Then, a subset of CpG sites was selected based upon the 7,136 genes found to be differentially expressed when vape users are compared to none vape users in the RNA-seq data. Finally, we corrected for epigenomic inflation using *bacon* [32] and corrected for multiple comparisons using FDR adjustment. Secondly, we conducted another targeted analysis which included only the CpG sites which were annotated to the genes found in the Gene Ontologies[33] for “inflammatory response”, “immune system process”, “immune response”, and “cilium”. This search also included CpG sites annotated to the *AHRR* gene, which has been the subject of tobacco-related research.[34] We again repeated the FDR p-value adjustment for only the CpG sites associated with these genes (FDR < 0.2).

**Assessment of differentially methylated regions (DMRs)**

The comb-p (combining p-values) method [35]was used and implemented in the python package comb*ined-pvalues 0.50.6* [36]. This method results in the identification of differentially methylated regions along with the CpG probes and genes which those regions overlap. Corrections for multiple testing in comb-p are made using the Sidak correction and all Sidak-corrected p-values up to a type-I error rate of 0.2 are reported.

* + - 1. **Results**
      2. ***Descriptive statistics***
      3. A total of 51 subjects participated in the study.  Vape users were characterized as adolescents who reported vaping within the past 6 months (n=13) while non-vape users were adolescents who did not have any vape exposure in the past 6 months (n=37). One subject did not report vape status and was therefore excluded from the study. The mean (standard deviation) age was 14.8 (1.4) years for vape users and 14.6 (1.4) years for non-vape users. We observed some demographic differences by vaping status. Most vape users were recruited in Pueblo (91%) and identified as LatinX (85%). 53% of subjects were female. Spirometry measurements were missing for all but one of the vape users and IOS data was available for most subjects (Table 1).
  1. **Table 1. Demographic characteristics and lung function testing results of study participants.** SD: standard deviation

|  |  |  |  |
| --- | --- | --- | --- |
|  | **Did Not Vape in Last 6 Months (non-vape users) (N=37)** | **Vaped in Last 6 Months (N=13) (vape users)** | **Total (N=50)** |
| **Sex** |  |  |  |
| Female | 17 (45.9%) | 8 (61.5%) | 25 (50.0%) |
| Male | 20 (54.1%) | 5 (38.5%) | 25 (50.0%) |
| **Age (years)** |  |  |  |
| Mean (SD) | 14.6 (1.4) | 14.8 (1.4) | 14.6 (1.4) |
| Range | 12.0 - 17.0 | 13.0 - 17.0 | 12.0 - 17.0 |
| **Recruitment Center** |  |  |  |
| Aurora | 15 (40.5%) | 0 (0.0%) | 15 (30.0%) |
| Commerce City/Denver | 13 (35.1%) | 1 (7.7%) | 14 (28.0%) |
| Pueblo | 9 (24.3%) | 12 (92.3%) | 21 (42.0%) |
| **Ethnicity** |  |  |  |
| LatinX | 23 (62.2%) | 11 (84.6%) | 34 (68.0%) |
| Non-LatinX | 14 (37.8%) | 2 (15.4%) | 16 (32.0%) |
| **R5** |  |  |  |
| N of Missing | 1 | 0 | 1 |
| Mean (SD) | 4.0 (0.9) | 5.0 (1.3) | 4.3 (1.1) |
| Range | 2.0 - 6.1 | 3.7 - 7.6 | 2.0 - 7.6 |
| **X20** |  |  |  |
| N of Missing | 4 | 2 | 6 |
| Mean (SD) | 0.1 (0.6) | 0.7 (0.9) | 0.2 (0.7) |
| Range | -1.1 - 2.4 | -1.0 - 2.3 | -1.1 - 2.4 |

***Lung function measures***

To assess the impact of vape exposure on measures of lung function, we tested the association of vape exposure with IOS measurements using two-sample t-tests of means. We observed significant differences in airway resistance by vaping status. Mean airway resistance (R) values calculated over a measurement period of 60 seconds at a frequency of 5 Hz (R5) revealed increased airway resistance in vaping (n=13) subjects compared to non-vaping control (n=37) subjects (p=0.026) [Fig 1], an early indication of increased airflow obstruction. Additionally, our results showed higher X20 (reactance) values in vape users compared to non-vape users (p=0.043), which suggests adverse effects of vaping on lung parenchyma.

1. **Chart, box and whisker chart

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2. **Fig 1. Differences in impulse oscillometry (IOS)** **measures between vape users and non-vape users.**

***Gene expression***

Nasal epithelial gene expression was utilized to assess the biological impact of vape exposure on the airway epithelium of the study subjects. Three participants were missing gene expression data and are excluded from subsequent analysis (N=47). 16,860 genes of the 60,651 genes were retained after removing low expressed genes. 7,136 were significantly differentially expressed between vaping subjects and control subjects after adjusting for age, sex, recruitment center, and two normalization factors (FDR<0.05) [37] (Figure 2). A total of 4,193 genes were underexpressed (e.g., *WNT5B*, *WNT3A, ACE2, IL7*) while 2,943 were overexpressed (e.g. *TNF*, *MUC5A, IL10, IL17C)* when vape users are compared to non-vape users (Table 2). Using a fold change cut of |log2(FC)| ≥ 2 there were 505 genes; 135 genes that were upregulated and 370 genes that were downregulated when comparing vape users with non-vape users (Supplementary Table 1).

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Table 2. The top twenty genes with the greatest fold-change (log2(FC)) when adolescents who vaped in the last 6 months (vape users) are compared to adolescents who did not vape (non-vape users).** The fold-change is the expression in vape users relative to the non-vape users (i.e., FC > 1 represents increased expression). | | | | | |
| **Ensembl ID** | **Gene Symbol** | **Log2(FC)** | **p-value** | **FDR** | **Associated function** |
| ***Negative log2 fold change*** | | | | | |
| ENSG00000147647 | *DPYS* | -3.9 | 5.8E-06 | 4.2E-05 | AGE-RAGE signaling pathway, oxidative stress |
| ENSG00000198838 | *RYR3* | -3.5 | 9.2E-04 | 3.5E-03 | 9+2 motile cilium |
| ENSG00000162782 | *TDRD5* | -3.4 | 1.1E-03 | 4.2E-03 | 9+2 motile cilium |
| ENSG00000152779 | *SLC16A12* | -3.4 | 8.7E-09 | 1.2E-07 | 9+0 motile cilium |
| ENSG00000039537 | *C6* | -3.4 | 4.6E-03 | 1.4E-02 | ADORA2B mediated anti-inflammatory cytokines production |
| ENSG00000280780 | *JAKMIP2-AS1* | -3.3 | 8.1E-12 | 2.6E-10 | Novel transcript |
| ENSG00000244067 | *GSTA2* | -3.3 | 1.7E-13 | 8.3E-12 | 9+0 motile cilium |
| ENSG00000260951 | *AC005100.1* | -3.2 | 1.5E-05 | 9.4E-05 | Novel transcript |
| ENSG00000277893 | *SRD5A2* | -3.1 | 1.4E-08 | 1.9E-07 | 9+0 motile cilium |
| ENSG00000268566 | *AC100781.1* | -3.1 | 6.5E-04 | 2.6E-03 | Novel transcript |
| ***Positive log2 fold change*** | | | | | |
| ENSG00000177257 | *DEFB4B* | 7.2 | 2.1E-03 | 7.3E-03 | ADORA2B mediated anti-inflammatory cytokines production |
| ENSG00000198692 | *EIF1AY* | 6.4 | 4.9E-03 | 1.5E-02 | 9+0 motile cilium |
| ENSG00000129824 | *RPS4Y1* | 6.4 | 4.6E-03 | 1.4E-02 | ADORA2B mediated anti-inflammatory cytokines production |
| ENSG00000012817 | *KDM5D* | 5.9 | 2.1E-03 | 7.1E-03 | 9+2 motile cilium |
| ENSG00000067048 | *DDX3Y* | 5.7 | 2.1E-04 | 9.6E-04 | 9+2 motile cilium |
| ENSG00000114374 | *USP9Y* | 4.8 | 5.8E-03 | 1.7E-02 | 9+0 motile cilium |
| ENSG00000232177 | *MTND4P24* | 4.7 | 4.2E-04 | 1.8E-03 | Novel transcript |
| ENSG00000225972 | *MTND1P23* | 4.2 | 9.3E-04 | 3.6E-03 | Novel transcript |
| ENSG00000201321 | *RNA5S9* | 3.7 | 3.8E-03 | 1.2E-02 | Novel transcript |
| ENSG00000183878 | *UTY* | 3.5 | 1.5E-04 | 7.1E-04 | 9+0 motile cilium |

***Enrichment analysis***

Pathways or ontologies were sourced from KEGG, Reactome and GO databases. The 505 Ensemble genes with |log2(FC)| ≥ 2 mapped to 476 Entrez gene IDs for enrichment analysis. Overall, vape users tended to have dysregulated expression of pathways associated with ciliogenesis and inflammation compared to the non-vape users (Table 2). We also observed the inflammatory response, immune system process, immune response, and cilium pathways and ontologies to be enriched (Tables 3-5).

**Chart

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**Figure 2 Differential gene expression results.** A volcano plot of differentially expressed genes between vape users and non-vape users (false discovery rate [FDR]<0.05), after adjusting for age, sex, recruitment center, and two normalization factors. Grey points (not significant) represent genes with an FDR > 0.5 and |Log2 FC| < 2.

**Table 3. Top 15 Gene Ontologies for differentially regulated genes when adolescents who vaped in the last 6 months (vape users) are compared to adolescents who did not vape (non-vape users).** Pathways with low false discovery rate (FDR) values and negative Normalized Enrichment Scores (NES) represent downregulated biological processes while those with low FDR and positive NES represent upregulated biological processes.



**Table 4. Top 15 Reactome pathways for differentially regulated genes when adolescents who vaped in the last 6 months (vape users) are compared to adolescents who did not vape (non-vape users).** Pathways with low false discovery rate (FDR) values and negative Normalized Enrichment Scores (NES) represent downregulated biological processes while those with low FDR and positive NES represent upregulated biological processes.



**Table 5. Top 15 KEGG pathways for differentially regulated genes when adolescents who vaped in the last 6 months (vape users) are compared to adolescents who did not vape (non-vape users).** Pathways with low false discovery rate (FDR) values and negative Normalized Enrichment Scores (NES) represent downregulated biological processes while those with low FDR and positive NES represent upregulated biological processes.

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***Sensitivity analysis***

With 92% (12/13) vape users and 42% 21/50 participants recruited at the Pueblo center, we sought to understand how this demographic imbalance acts as a potential confounder or source of multicollinearity. As such we restricted analysis to participants recruited from the Pueblo center and compared it to the overall model containing all study participants to assess differences in estimates and significant gene composition between the models. The top 2,000 significant genes for the restricted and full models are identical both before and after the selected cutoff value (|Log2(FC) | > 2). The variation of Log2(FC) from the top 2,000 genes in the model with only subjects and the model with only Pueblo subjects were minimal (Supplementary Figure 2, Supplementary Table 2).

***Targeted DNA Methylation Analysis***

One CpG site, cg02123174, was significantly differentially methylated at a type-I error rate of 0.05. The site maps to *REXO1* and was less methylated in vape users compared to non-vape users. This result is consistent with RNA-Seq results where the gene was up-regulated (0.396, FDR = 0.02). One additional site, cg11903190 was marginally significant at an FDR-adjusted p-value of 0.06 and maps to *CERK*, which was also up-regulated in the RNA-Seq results (0.395, FDR = 0.002). At a type-I error rate of 0.2, there were 36 differentially methylated sites. Supplementary Table 3 gives the top 10 sorted by adjusted p-value and estimate size while Supplementary Table 3 shows boxplots of five of such CpG sites (and their associated genes).

***Identification of differentially methylated regions (DMRs)***

Supplementary Table 4 shows all DMRs with Sidak-adjusted p-values < 0.1 in addition to their associated genes from the UCSC genome browser. Four CpG sites: cg13300473;cg15775218;cg15913725;cg25411977, which are associated with *EIPR1* (a gene identified in the ciliary landscape super path [38]), was located in a DMR that was highly statistically significant (Sidak-adjusted p value <0.001). Likewise, cg02123174, which is associated with *REXO1* was less methylated (Sidak-adjusted p value = 0.037).

**Discussion**

Our results suggest that adolescent vape exposure is associated with increased airflow obstruction and increased expression of inflammatory genes in the nasal epithelium of vape users. Our work provides critical evidence of the negative impact of vape exposure on respiratory outcomes even in early adolescence. To the best of our knowledge, these data are the first to demonstrate that vape exposure is associated with abnormal lung function in early adolescence and shows the detrimental impact of vape exposure during a critical window of lung function development.

Our findings are consistent with existing literature in adults demonstrating that short-term exposure to e-cigs increases airway resistance and is associated with decreased measures of lung function after exposure [12]. We identified significant differences in IOS measures between vape users and non-vape users. To date, there have been few studies in humans looking at e-cig use and lung function measures and none have looked at long term exposures and their effects on lung function. A previous study on the acute effects of vape exposure on airway physiology and respiratory symptoms in COPD smokers, asthmatic smokers, "healthy" smokers and healthy never smokers after e-cig use identified an increase in airways resistance in asthmatic (p=0.034) and healthy smokers (p=0.004). Additionally, an increase in airway resistance was noted in never smokers after using e-cigs with (p<0.005) or without (p<0.001) nicotine [39]. Another study on documented e-cig exposures revealed adverse changes to respiratory metrics and markers of inflammation after a 5 minute e-cig use session. The authors observed that R5, a marker of total airway resistance, had increased post e-cig use in both healthy controls and mild asthmatics. Notably, the increase in airway resistance was significantly higher among the mild asthmatics [40].

A 2022 systematic analysis of the effects of e-cigs on lung function when compared to traditional cigarettes found that there were statistically significant increases in airflow resistance (Z5, R5 and R10), after e-cig inhalation among asthmatic smokers using data from Europe between 2018–2020 [41]. This systematic analysis used data from n= 10 to 408 participants (mean age ranging from 22.6 – 58 years) with most of the studies (16/18) assessing lung function after about 5 min to 1 month of e-cig use. The impacts of e-cig use on airway physiology have also been documented during passive vape exposures. In a study with 15 subjects with repeated measures of lung function during passive vape exposure, there were no significant changes in FEV1/FVC when exposed to one hour of passive e-cigarette smoking (indicative: 2.3% reduction in FEV1/FVC) [42]. A cross-over study also looked at 40 healthy nonsmokers (18–35 years old) exposed to e-cig emissions produced at two resistance settings, 0.5 ohm and 1.5 ohm. At the 1.5 ohm session, R5 showed a post exposure decrease trend that was not significant (0.39 pre to 0.38 kPa/L/s post exposure) [43].

While these studies are short term in nature, we believe our data which assessed lung function at about 6 months of e-cig use adds to the current state of the science on the adverse impacts on the small airways after e-cig exposure. Further studies are needed to understand the respiratory health impacts of long-term e-cig use. Additionally, these previous studies were in adult populations, some of whom had comorbidities such as asthma or chronic obstructive pulmonary disease. Our study population was in relatively healthy adolescents without previously reported respiratory disease and is the first study to our knowledge that documents the adverse impact of e-cig use on lung function.

Furthermore, we demonstrate differences in nasal epithelial gene expression in vape users compared to non-vape users. Importantly, the top four genes (*CEACAM4, MMP25, NCF1* and *NFAM1)* that were overexpressed in vape users compared to non-vape users, were associated with inflammatory processes or in immune function (Fig 3). For instance, *CEACAM4* is an orphan receptor of the *CEACAM* family that is associated with phagocytic function [44]. *MMP25* is a metalloprotease that regulates innate immune response through NF-κB signaling [45]. Mutations in *NCF1* is associated with chronic granulomatous disease [46] and variants of this gene are associated with an increased risk of developing autoimmune diseases [47-49] and recurrent spontaneous abortion [50]. *NFAM1* promotes pro-inflammatory cytokine production in both mouse and human monocytes and has been identified as a potential therapeutic target for treatment of autoimmune disease [51]which are characterized by abnormal inflammatory response [52].

Three of the top genes (*CAPS, DNAH7* and  *NWD1)* that were downregulated among vape users, compared to non-vape users have significant implications for respiratory health based on reports in the literature. *CAPS* encodes the calcium binding protein calcyphosine which is associated with cellular proliferation and differentiation [53]. *DNAH7* is associated with ciliary dysfunction and may be important in understanding the molecular pathogenesis of Middle East respiratory syndrome coronavirus (MERS-CoV) [54] and COVID-19 infections [55]. *NWD1* has been demonstrated to modulate androgen receptor protein levels [56] and is involved in axonal growth, with high protein levels detected in the brain of a mouse model for seizure and temporal lobe epilepsy [57]. A recent study demonstrated that the knockdown of *NWD1* inhibited dendritic growth and synaptogenesis [58]. Taken together, vaping may be associated with increased risk of inflammation and decreased ciliogenesis thereby impacting airflow obstruction.

Our results are supported by previously published investigations on the impact of e-cig use. In a cohort of ten healthy never smokers, short term e-cig use was associated with altered transcriptomes of small airway epithelial cells and alveolar macrophages among all subjects [59]. Interestingly, two genes identified in this short-term study were dysregulated in the opposite direction in our study. *NDC80* (FC: 0.56) and *PTGER3* (FC: -1.3) had differential expression in directions opposite to our current study. However, differences in e-cig products, demographics of the study population, duration of exposure as well as the location of the biological sample (brushing nasal epithelial vs brushing the 10th–12th order bronchi) limits direct comparisons with our work. Further studies are needed to understand the effects of the increasing prevalence of e-cig use. Indeed, Sayed et al compared sputum and salivafrom e-cig users and nonusers and observed reductions in markers of airway inflammation among e-cig users compared to nonusers [60]. However, plasma concentrations of certain inflammatorycytokines, chemokines, and growth factors were higher among e-cig users. Hence it is possible that changes in airway inflammatory markers may be a counter-response to general inflammation caused by chronic e-cig use [60] and additional studies are warranted.

Several members of the mucin family (eg *MUC5AC, MUC12*) were differentially expressed among vape users and nonvape users. Mucins are O-glycosylated proteins that play an essential role in forming protective mucous barriers on epithelial surfaces and have been implicated in epithelial renewal and differentiation. These glycoproteins also play a role in intracellular signaling. In our study, several signaling pathways such as ‘cytokine-mediated’, ‘cell surface receptor’, ‘signal transduction’, ‘cytokine signaling in immune system’, ‘G alpha signaling events’, ‘chemokine signaling, ‘NOD-like receptor’ and ‘B cell receptor’ signaling pathways were enriched. Another noteworthy pathway recently implicated in developmental biology is the *wnt* signaling pathway. In our study, *WNT5B, WNT3A* and *WNT4* were underexpressed in vape users compared to nonvape users. W*nt* signaling has been recently highlighted as a pathway whose dysregulation can affect lung disease development later in life [61, 62].

Pathways enriched for ciliary function involved genes such as the DNAH family. Of note, of the 12 members of the DNAH family of genes that were differentially expressed in our data set, 11 were underexpressed in vape users. *DNAH17* was the only gene that was overexpressed*.* Most of the 11 DNAH genes have been detected in the lung and have been implicated in ciliary dyskinesia in airway epithelial cells [63] and DNAH17 is mostly expressed in the testis and are less abundant in the lung [64]. Since e-cig use is associated with significant inflammation in the lungs, albeit the dysregulation in the immune landscape differs from that of tobacco use, it is important for future studies to focus on e-cig use and its impact on systemic function as well [65].

An interesting pathway in our enrichment analysis was the inflammasome pathway. *IL1B* was significantly upregulated in vape users (FC: 2.1; FDR < 0.001) and this has been suggested that inflammasomes are activated, and like Lee et al, *CXCL1, CXCL2* and *NOD2* were upregulated in vape users [65]. Inflammasomes (such as nucleotide-binding oligomerization domain, leucine rich repeat and pyrin domain containing (NLRP)) are large protein structures primarily located in macrophages that respond to inflammatory signals [65]. Upon activation, inflammasomes cleave pro-IL1B and pro-*IL18* into *IL1B* and *IL18*, to signal the presence of xenobiotics and initiate the inflammatory response [66]. Several members of the NLRP family were differentially expressed when vape users were compared to non-vape users (*NLRP1, NLRP3, NLRP6, NLRP12,* and *NLRP14*). All except *NLRP14* were upregulated. The dysregulation of inflammasome pathway may be an important step in immune dysregulation during chronic e-cig use. For instance, an inflammatory cytokine such as *IL1B* (upregulated in vape users) can stimulate the production of other inflammatory cytokines (eg *IL6* and *TNF*) which were also upregulated in vape users vs non-vape users in our study. Future studies may need to assess the types of e-cig devices that may stimulate different pathways due to chronic e-cig use. The study by Hickman et al highlights the importance of assessing device type. Data from third- and fourth-generation e-cig users and found an overall suppression of host defense associated with fourth-generation e-cig use [67]. This may be due to new/emerging formulations in the e-cigs such as nicotine salts and future studies need to be designed to understand the health consequences of emerging e-cigs [68].

Finally, our results suggest that DNA methylation may be one of the mechanisms by which vaping exerts its effects on the lungs. *REXO1* and *CERK* were differentially methylated when vape users are compared to non-vape users. While not much is known about REXO1, its over expression has been associated with cell proliferation, migration, and invasion in cervical cancer [69]. Notably, *REXO1* was in a significantly methylated region in our study and a recent study by Herrera-Luis et al among Puerto Rican and Mexican American children and young adults (average age 12-14 years) with asthma, identified REXO1 in one of two differentially methylated regions as associated with pre-FEV1/FVC [70] Interestingly, the association signals for the Herrera-Luis et al study were enriched for inflammatory processes and the authors concluded that REXO1 is associated with airflow limitation in children.

*CERK* can be activated by *IL1B*, and its activity has been observed in neutrophils and the lung epithelium [71, 72]. The gene encodes ceramide kinase and has been implicated as an important regulatory component of inflammatory response [73]. Notably, ceramide kinase converts ceramide to cermide-1-phosphate [74], which plays a role in linking Prostaglandin E2, neurotransmitters, and airway epithelial inflammation [73]. Together, our data provide further insights into the mechanisms involved in adverse impacts on lung function.

A pediatric pulmonary review concluded that vaping may increase risk of developing chronic lung disease [75]. The authors indicated that vaping is associated with both in vitro and in vivo airway mucociliary dysfunction, increased epithelial cell, macrophage death, as well as dysregulation of airway epithelium. Indeed, data from our current study point to these adverse respiratory impacts among adolescent vape users. Our findings demonstrate that significant changes in airway resistance that result from vape exposure may precede the development of adverse respiratory symptoms and that vape exposure negatively impacts adolescent lung function during a critical stage of their lung development. We also establish that vape exposure results in changes in gene expression in inflammatory pathways and genes involved in ciliary function in the nasal epithelium and that this may be biological mechanism that underlies the development of airflow obstruction in this at-risk population.

Limitations of our study include the small sample size, the cross-sectional nature of the study, and the lack of exposure assessment specific to e-cigarettes in this cohort. Future investigations in this at-risk adolescent population will be needed to assess the longitudinal impact of habitual vape use on lung function outcomes and should include a more comprehensive exposure assessment of the contents of vape aerosols. Our data indicates that vaping is associated with impaired lung function in adolescents and extensive changes in nasal epithelial gene expression. While limited in sample size, our results add to the currently limited knowledge on the long-term effects of vape use. Given the paucity of information on the effects of vaping on the airway epithelium and the high-risk youth population with access to these devices, our work suggests that further research is needed. Such future work will help characterize chronic vape use and its impact on lung function, nasal epithelial gene expression, and how vaping cessation may reverse these changes.

**References**

1. Rowell, T.R. and R. Tarran, Will chronic e-cigarette use cause lung disease? *Am J Physiol Lung Cell Mol Physiol*, **2015**. 309(12): p. L1398-409.

2. National Academies of Sciences, E., Medicine, Health, D. Medicine, H. Board on Population, P. Public Health, and S. Committee on the Review of the Health Effects of Electronic Nicotine Delivery, *Public Health Consequences of E-Cigarettes*, in *Public Health Consequences of E-Cigarettes*, D.L. Eaton, L.Y. Kwan, and K. Stratton, Editors. 2018, National Academies Press (US) Copyright 2018 by the National Academy of Sciences. All rights reserved.: Washington (DC).

3. Walley, S.C. and B.P. Jenssen, Electronic Nicotine Delivery Systems*.* *Pediatrics*, **2015**. 136(5): p. 1018-26.

4. Douglass, B., S. Solecki, and T. Fay-Hillier, The Harmful Consequences of Vaping: A Public Health Threat*.* *J Addict Nurs*, **2020**. 31(2): p. 79-84.

5. DHHS, U.S. Department of Health & Human Services (2016). E-cigarette use among youth and young adults: A report of the surgeon general—Executive summary. U.S. Department of Health and Human Services Centers for Disease Control and Prevention, National Center for Chronic Disease Prevention and Health Promotion, Office on Smoking and Health. <https://e-cigarettes.surgeongeneral.gov/documents/2016_sgr_full_report_non-508.pdf>*.* **2016**.

6. Walley, S.C., K.M. Wilson, J.P. Winickoff, and J. Groner, A Public Health Crisis: Electronic Cigarettes, Vape, and JUUL*.* *Pediatrics*, **2019**. 143(6).

7. Hammig, B., P. Daniel-Dobbs, and H. Blunt-Vinti, Electronic cigarette initiation among minority youth in the United States*.* *Am J Drug Alcohol Abuse*, **2017**. 43(3): p. 306-310.

8. Cullen  KA, A.B., Gentzke  AS, Apelberg  BJ, Jamal  A, King  BA, Notes from the field: use of electronic cigarettes and any tobacco product among middle and high school students—United States, 2011-2018.  MMWR Morb Mortal Wkly Rep; 67(45):1276-1277. **2018**.

9. Miech, R., L. Johnston, P.M. O'Malley, J.G. Bachman, and M.E. Patrick, Adolescent Vaping and Nicotine Use in 2017-2018 - U.S. National Estimates*.* *N Engl J Med*, **2019**. 380(2): p. 192-193.

10. Miech, R., A.M. Leventhal, P.M. O'Malley, L.D. Johnston, and J.L. Barrington-Trimis, Failed Attempts to Quit Combustible Cigarettes and e-Cigarettes Among US Adolescents*.* *Jama*, **2022**. 327(12): p. 1179-1181.

11. Lee, J., V. Taneja, and R. Vassallo, Cigarette smoking and inflammation: cellular and molecular mechanisms*.* *J Dent Res*, **2012**. 91(2): p. 142-9.

12. Chun, L.F., F. Moazed, C.S. Calfee, M.A. Matthay, and J.E. Gotts, Pulmonary toxicity of e-cigarettes*.* *American journal of physiology. Lung cellular and molecular physiology*, **2017**. 313(2): p. L193-L206.

13. Suzuki, T., K. Tsushima, N. Kawata, T. Matsumura, Y. Matsuura, Y. Ichimura, J. Terada, S. Sakao, Y. Tada, N. Tanabe, and K. Tatsumi, Estimation using the impulse oscillation system in patients with pulmonary sarcoidosis*.* *Sarcoidosis Vasc Diffuse Lung Dis*, **2015**. 32(2): p. 144-50.

14. Miller, M.R., R. Crapo, J. Hankinson, V. Brusasco, F. Burgos, R. Casaburi, A. Coates, P. Enright, C.P. van der Grinten, P. Gustafsson, R. Jensen, D.C. Johnson, N. MacIntyre, R. McKay, D. Navajas, O.F. Pedersen, R. Pellegrino, G. Viegi, and J. Wanger, General considerations for lung function testing*.* *Eur Respir J*, **2005**. 26(1): p. 153-61.

15. Wong, A., K. Hardaker, P. Field, J. Huvanandana, G.G. King, H. Reddel, H. Selvadurai, C. Thamrin, and P.D. Robinson, Home-based Forced Oscillation Technique Day-to-Day Variability in Pediatric Asthma*.* *Am J Respir Crit Care Med*, **2019**. 199(9): p. 1156-1160.

16. Culver, B.H., B.L. Graham, A.L. Coates, J. Wanger, C.E. Berry, P.K. Clarke, T.S. Hallstrand, J.L. Hankinson, D.A. Kaminsky, N.R. MacIntyre, M.C. McCormack, M. Rosenfeld, S. Stanojevic, D.J. Weiner, and A.T.S.C.o.P.S.f.P.F. Laboratories, Recommendations for a Standardized Pulmonary Function Report. An Official American Thoracic Society Technical Statement*.* *Am J Respir Crit Care Med*, **2017**. 196(11): p. 1463-1472.

17. Lundblad, L.K.A., R. Miletic, E. Piitulainen, and P. Wollmer, Oscillometry in Chronic Obstructive Lung Disease: In vitro and in vivo evaluation of the impulse oscillometry and tremoflo devices*.* *Sci Rep*, **2019**. 9(1): p. 11618.

18. Ward, C.M., T.H. To, and S.M. Pederson, ngsReports: a Bioconductor package for managing FastQC reports and other NGS related log files*.* *Bioinformatics*, **2020**. 36(8): p. 2587-2588.

19. He, B., R. Zhu, H. Yang, Q. Lu, W. Wang, L. Song, X. Sun, G. Zhang, S. Li, J. Yang, G. Tian, P. Bing, and J. Lang, Assessing the Impact of Data Preprocessing on Analyzing Next Generation Sequencing Data*.* *Front Bioeng Biotechnol*, **2020**. 8: p. 817.

20. Frankish, A., M. Diekhans, A.M. Ferreira, R. Johnson, I. Jungreis, J. Loveland, J.M. Mudge, C. Sisu, J. Wright, J. Armstrong, I. Barnes, A. Berry, A. Bignell, S. Carbonell Sala, J. Chrast, F. Cunningham, T. Di Domenico, S. Donaldson, I.T. Fiddes, C. Garcia Giron, J.M. Gonzalez, T. Grego, M. Hardy, T. Hourlier, T. Hunt, O.G. Izuogu, J. Lagarde, F.J. Martin, L. Martinez, S. Mohanan, P. Muir, F.C.P. Navarro, A. Parker, B. Pei, F. Pozo, M. Ruffier, B.M. Schmitt, E. Stapleton, M.M. Suner, I. Sycheva, B. Uszczynska-Ratajczak, J. Xu, A. Yates, D. Zerbino, Y. Zhang, B. Aken, J.S. Choudhary, M. Gerstein, R. Guigo, T.J.P. Hubbard, M. Kellis, B. Paten, A. Reymond, M.L. Tress, and P. Flicek, GENCODE reference annotation for the human and mouse genomes*.* *Nucleic Acids Res*, **2019**. 47(D1): p. D766-D773.

21. Dobin, A., C.A. Davis, F. Schlesinger, J. Drenkow, C. Zaleski, S. Jha, P. Batut, M. Chaisson, and T.R. Gingeras, STAR: ultrafast universal RNA-seq aligner*.* *Bioinformatics*, **2013**. 29(1): p. 15-21.

22. Wieczorek D, Stayer C, and Schagat T, Automated DNA Purification from Oragene•DNA/Saliva Samples Using the Maxwell® 16 System. Promega Corporation Web site. <http://www.promega.com/resources/pubhub/enotes/automated-dna-purification-from-oragene-dna-saliva-samples-using-the-maxwell-16-system/> Updated 2008. Accessed September 5 2017 **2008**.

23. Risso, D., J. Ngai, T.P. Speed, and S. Dudoit, Normalization of RNA-seq data using factor analysis of control genes or samples*.* *Nat Biotechnol*, **2014**. 32(9): p. 896-902.

24. Robinson, M.D., D.J. McCarthy, and G.K. Smyth, edgeR: a Bioconductor package for differential expression analysis of digital gene expression data*.* *Bioinformatics*, **2010**. 26(1): p. 139-40.

25. Love, M.I., W. Huber, and S. Anders, Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2*.* *Genome Biol*, **2014**. 15(12): p. 550.

26. Korotkevich, G., V. Sukhov, N. Budin, B. Shpak, M.N. Artyomov, and A. Sergushichev, Fast gene set enrichment analysis*.* *bioRxiv*, **2021**: p. 060012.

27. Xiao, Y., T.H. Hsiao, U. Suresh, H.I. Chen, X. Wu, S.E. Wolf, and Y. Chen, A novel significance score for gene selection and ranking*.* *Bioinformatics*, **2014**. 30(6): p. 801-7.

28. Korotkevich, G., V. Sukhov, N. Budin, B. Shpak, M.N. Artyomov, and A.J.B. Sergushichev, Fast gene set enrichment analysis*.* **2016**: p. 060012.

29. Durinck, S., P.T. Spellman, E. Birney, and W. Huber, Mapping identifiers for the integration of genomic datasets with the R/Bioconductor package biomaRt*.* *Nat Protoc*, **2009**. 4(8): p. 1184-91.

30. Durinck, S., Y. Moreau, A. Kasprzyk, S. Davis, B. De Moor, A. Brazma, and W. Huber, BioMart and Bioconductor: a powerful link between biological databases and microarray data analysis*.* *Bioinformatics*, **2005**. 21(16): p. 3439-40.

31. Hansen KD. IlluminaHumanMethylationEPICanno.ilm10b4.hg19: Annotation for Illumina’s EPIC Methylation Arrays.; 2017. <https://bitbucket.com/kasperdanielhansen/Illumina_EPIC>.

32. van Iterson, M., E.W. van Zwet, and B.T. Heijmans, Controlling bias and inflation in epigenome- and transcriptome-wide association studies using the empirical null distribution*.* *Genome Biol*, **2017**. 18(1): p. 19.

33. The Gene Ontology resource: enriching a GOld mine*.* *Nucleic Acids Res*, **2021**. 49(D1): p. D325-d334.

34. Richmond, R.C., C. Sillero-Rejon, J.N. Khouja, C. Prince, A. Board, G. Sharp, M. Suderman, C.L. Relton, M. Munafò, and S.H. Gage, Investigating the DNA methylation profile of e-cigarette use*.* *Clin Epigenetics*, **2021**. 13(1): p. 183.

35. Kechris, K.J., B. Biehs, and T.B. Kornberg, Generalizing moving averages for tiling arrays using combined p-value statistics*.* *Stat Appl Genet Mol Biol*, **2010**. 9(1): p. Article29.

36. Pedersen, B.S., D.A. Schwartz, I.V. Yang, and K.J. Kechris, Comb-p: software for combining, analyzing, grouping and correcting spatially correlated P-values*.* *Bioinformatics*, **2012**. 28(22): p. 2986-8.

37. Risso, D., J. Ngai, T.P. Speed, and S. Dudoit, Normalization of RNA-seq data using factor analysis of control genes or samples*.* *Nature biotechnology*, **2014**. 32(9): p. 896-902.

38. Weizmann Institute of Science. *Ciliary landscape Singleton SuperPath*. 2023 3/17/2023]; <https://pathcards.genecards.org/card/ciliary_landscape>]. Available from: <https://pathcards.genecards.org/card/ciliary_landscape>.

39. Palamidas, A., S. Tsikrika, P.A. Katsaounou, S. Vakali, S.A. Gennimata, G. Kaltsakas, C. Gratziou, and N. Koulouris, Acute effects of short term use of ecigarettes on Airways Physiology and Respiratory Symptoms in Smokers with and without Airway Obstructive Diseases and in Healthy non smokers*.* *Tob Prev Cessat*, **2017**. 3: p. 5.

40. Lappas, A.S., A.S. Tzortzi, E.M. Konstantinidi, S.I. Teloniatis, C.K. Tzavara, S.A. Gennimata, N.G. Koulouris, and P.K. Behrakis, Short-term respiratory effects of e-cigarettes in healthy individuals and smokers with asthma*.* *Respirology*, **2018**. 23(3): p. 291-297.

41. Song, Y., X. Li, C. Li, S. Xu, Y. Liu, and X. Wu, What Are the Effects of Electronic Cigarettes on Lung Function Compared to Non-Electronic Cigarettes? A Systematic Analysis*.* *Int J Public Health*, **2022**. 67: p. 1604989.

42. Flouris, A.D., M.S. Chorti, K.P. Poulianiti, A.Z. Jamurtas, K. Kostikas, M.N. Tzatzarakis, A. Wallace Hayes, A.M. Tsatsakis, and Y. Koutedakis, Acute impact of active and passive electronic cigarette smoking on serum cotinine and lung function*.* *Inhal Toxicol*, **2013**. 25(2): p. 91-101.

43. Tzortzi, A., S.I. Teloniatis, G. Matiampa, G. Bakelas, V.K. Vyzikidou, C. Vardavas, P.K. Behrakis, and E. Fernandez, Passive exposure to e-cigarette emissions: Immediate respiratory effects*.* *Tob Prev Cessat*, **2018**. 4: p. 18.

44. Delgado Tascón, J., J. Adrian, K. Kopp, P. Scholz, M.P. Tschan, K. Kuespert, and C.R. Hauck, The granulocyte orphan receptor CEACAM4 is able to trigger phagocytosis of bacteria*.* *J Leukoc Biol*, **2015**. 97(3): p. 521-31.

45. Soria-Valles, C., A. Gutiérrez-Fernández, F.G. Osorio, D. Carrero, A.A. Ferrando, E. Colado, M.S. Fernández-García, E. Bonzon-Kulichenko, J. Vázquez, A. Fueyo, and C. López-Otín, MMP-25 Metalloprotease Regulates Innate Immune Response through NF-κB Signaling*.* *J Immunol*, **2016**. 197(1): p. 296-302.

46. Chung, A.G., M.M. Cyr, and A.K. Ellis, Newly diagnosed chronic granulomatous disease in a 44 year old male presenting with recurrent groin cellulitis and colitis*.* *Allergy Asthma Clin Immunol*, **2013**. 9(1): p. 9.

47. Meng, Y., J. Ma, C. Yao, Z. Ye, H. Ding, C. Liu, J. Li, G. Li, Y. He, J. Li, Z. Yin, L. Wu, H. Zhou, and N. Shen, The NCF1 variant p.R90H aggravates autoimmunity by facilitating the activation of plasmacytoid dendritic cells*.* *The Journal of clinical investigation*, **2022**. 132(16): p. e153619.

48. Zhong, J., A.C.Y. Yau, and R. Holmdahl, Independent and inter-dependent immunoregulatory effects of NCF1 and NOS2 in experimental autoimmune encephalomyelitis*.* *Journal of neuroinflammation*, **2020**. 17(1): p. 113-113.

49. Zhang, L., J. Wax, R. Huang, F. Petersen, and X. Yu, Meta-Analysis and Systematic Review of the Association between a Hypoactive NCF1 Variant and Various Autoimmune Diseases*.* *Antioxidants (Basel, Switzerland)*, **2022**. 11(8): p. 1589.

50. Du, M., H. Gu, Y. Li, L. Huang, M. Gao, H. Xu, H. Deng, W. Zhong, X. Liu, and X. Zhong, A missense variant in NCF1 is associated with susceptibility to unexplained recurrent spontaneous abortion*.* *Open life sciences*, **2022**. 17(1): p. 1443-1450.

51. Juchem, K.W., A.P. Gounder, J.P. Gao, E. Seccareccia, N. Yeddula, N.J. Huffmaster, A. Côté-Martin, S.E. Fogal, D. Souza, S.S. Wang, E.R.A. Glynn, I. Yung, J. Ritchie, L. Li, J. Zheng, M.L. Mbow, J. Li, and S.K. Chanda, NFAM1 Promotes Pro-Inflammatory Cytokine Production in Mouse and Human Monocytes*.* *Front Immunol*, **2021**. 12: p. 773445.

52. Duan, L., X. Rao, and K.R. Sigdel, Regulation of Inflammation in Autoimmune Disease*.* *J Immunol Res*, **2019**. 2019: p. 7403796.

53. Clément, S., J.E. Dumont, and S. Schurmans, Loss of calcyphosine gene expression in mouse and other rodents*.* *Biochem Biophys Res Commun*, **1997**. 232(2): p. 407-13.

54. Lado, S., J.P. Elbers, M. Plasil, T. Loney, P. Weidinger, J.V. Camp, J. Kolodziejek, J. Futas, D.A. Kannan, P. Orozco-terWengel, P. Horin, N. Nowotny, and P.A. Burger, Innate and Adaptive Immune Genes Associated with MERS-CoV Infection in Dromedaries*.* *Cells*, **2021**. 10(6).

55. Hu, J., C. Li, S. Wang, T. Li, and H. Zhang, Genetic variants are identified to increase risk of COVID-19 related mortality from UK Biobank data*.* *medRxiv*, **2020**.

56. Correa, R.G., M. Krajewska, C.F. Ware, M. Gerlic, and J.C. Reed, The NLR-related protein NWD1 is associated with prostate cancer and modulates androgen receptor signaling*.* *Oncotarget*, **2014**. 5(6): p. 1666-82.

57. Yang, Q., Z. Huang, Y. Luo, F. Zheng, Y. Hu, H. Liu, S. Zhu, M. He, D. Xu, Y. Li, M. Yang, Y. Yang, X. Wei, X. Gao, W. Wang, J. Ma, Y. Ma, X. Wang, and Q. Wang, Inhibition of Nwd1 activity attenuates neuronal hyperexcitability and GluN2B phosphorylation in the hippocampus*.* *EBioMedicine*, **2019**. 47: p. 470-483.

58. Wu, Y., Q. Fu, X. Huang, Y. Luo, S. Wan, M. Peng, S. Su, X. Xu, Y. Li, X. Li, D. Sun, and C. Ke, NWD1 facilitates synaptic transmission and contributes to neuropathic pain*.* *Neuropharmacology*, **2022**. 205: p. 108919.

59. Staudt, M.R., J. Salit, R.J. Kaner, C. Hollmann, and R.G. Crystal, Altered lung biology of healthy never smokers following acute inhalation of E-cigarettes*.* *Respir Res*, **2018**. 19(1): p. 78.

60. Sayed, I.M., J.A. Masso-Silva, A. Mittal, A. Patel, E. Lin, A. Moshensky, J. Shin, C.M. Bojanowski, S. Das, P. Akuthota, and L.E. Crotty Alexander, Inflammatory phenotype modulation in the respiratory tract and systemic circulation of e-cigarette users: a pilot study*.* *Am J Physiol Lung Cell Mol Physiol*, **2021**. 321(6): p. L1134-l1146.

61. Cahill, K.M., M.R. Gartia, S. Sahu, S.R. Bergeron, L.M. Heffernan, D.B. Paulsen, A.L. Penn, and A. Noël, In utero exposure to electronic-cigarette aerosols decreases lung fibrillar collagen content, increases Newtonian resistance and induces sex-specific molecular signatures in neonatal mice*.* *Toxicological Research*, **2022**. 38(2): p. 205-224.

62. Cahill, K.M., T.K. Johnson, Z. Perveen, M. Schexnayder, R. Xiao, L.M. Heffernan, I.M. Langohr, D.B. Paulsen, A.L. Penn, and A. Noël, In utero exposures to mint-flavored JUUL aerosol impair lung development and aggravate house dust mite-induced asthma in adult offspring mice*.* *Toxicology*, **2022**. 477: p. 153272.

63. Imtiaz, F., R. Allam, K. Ramzan, and M. Al-Sayed, Variation in DNAH1 may contribute to primary ciliary dyskinesia*.* *BMC Med Genet*, **2015**. 16: p. 14.

64. Whitfield, M., L. Thomas, E. Bequignon, A. Schmitt, L. Stouvenel, G. Montantin, S. Tissier, P. Duquesnoy, B. Copin, S. Chantot, F. Dastot, C. Faucon, A.L. Barbotin, A. Loyens, J.P. Siffroi, J.F. Papon, E. Escudier, S. Amselem, V. Mitchell, A. Touré, and M. Legendre, Mutations in DNAH17, Encoding a Sperm-Specific Axonemal Outer Dynein Arm Heavy Chain, Cause Isolated Male Infertility Due to Asthenozoospermia*.* *Am J Hum Genet*, **2019**. 105(1): p. 198-212.

65. Lee, A.C., J. Chakladar, W.T. Li, C. Chen, E.Y. Chang, J. Wang-Rodriguez, and W.M. Ongkeko, Tobacco, but Not Nicotine and Flavor-Less Electronic Cigarettes, Induces ACE2 and Immune Dysregulation*.* *Int J Mol Sci*, **2020**. 21(15).

66. Lu, A. and H. Wu, Structural mechanisms of inflammasome assembly*.* *Febs j*, **2015**. 282(3): p. 435-44.

67. Hickman, E., A. Payton, P. Duffney, H. Wells, A.S. Ceppe, S. Brocke, A. Bailey, M.E. Rebuli, C. Robinette, B. Ring, J.E. Rager, N.E. Alexis, and I. Jaspers, Biomarkers of Airway Immune Homeostasis Differ Significantly with Generation of E-Cigarettes*.* *Am J Respir Crit Care Med*, **2022**. 206(10): p. 1248-1258.

68. Sullivan, L. and L.E. Crotty Alexander, A Problem for Generations: Impact of E-Cigarette Type on Immune Homeostasis*.* *Am J Respir Crit Care Med*, **2022**. 206(10): p. 1195-1197.

69. Zhang, Y., X. Li, J. Zhang, and L. Mao, Circ-CCDC66 upregulates REXO1 expression to aggravate cervical cancer progression via restraining miR-452-5p*.* *Cancer Cell Int*, **2021**. 21(1): p. 20.

70. Herrera-Luis, E., A. Li, A.C.Y. Mak, J. Perez-Garcia, J.R. Elhawary, S.S. Oh, D. Hu, C. Eng, K.L. Keys, S. Huntsman, K.B. Beckman, L.N. Borrell, J. Rodriguez-Santana, E.G. Burchard, and M. Pino-Yanes, Epigenome-wide association study of lung function in Latino children and youth with asthma*.* *Clin Epigenetics*, **2022**. 14(1): p. 9.

71. Hinkovska-Galcheva, V.T., L.A. Boxer, P.J. Mansfield, D. Harsh, A. Blackwood, and J.A. Shayman, The formation of ceramide-1-phosphate during neutrophil phagocytosis and its role in liposome fusion*.* *J Biol Chem*, **1998**. 273(50): p. 33203-9.

72. Rile, G., Y. Yatomi, T. Takafuta, and Y. Ozaki, Ceramide 1-phosphate formation in neutrophils*.* *Acta Haematol*, **2003**. 109(2): p. 76-83.

73. Pettus, B.J., A. Bielawska, S. Spiegel, P. Roddy, Y.A. Hannun, and C.E. Chalfant, Ceramide kinase mediates cytokine- and calcium ionophore-induced arachidonic acid release*.* *J Biol Chem*, **2003**. 278(40): p. 38206-13.

74. Hoeferlin, L.A., D.S. Wijesinghe, and C.E. Chalfant, The role of ceramide-1-phosphate in biological functions*.* *Handb Exp Pharmacol*, **2013**(215): p. 153-66.

75. Kaslow, J.A., C. Rosas-Salazar, and P.E. Moore, E-cigarette and vaping product use-associated lung injury in the pediatric population: A critical review of the current literature*.* *Pediatr Pulmonol*, **2021**. 56(7): p. 1857-1867.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Supplementary Table 1: All transcripts with the at least 2 fold-change (log2FC) when adolescents who vaped in the last 6 months (vape users) are compared to adolescents who did not vape (non-vape users). The fold-change is the expression in the vaping group relative to the control group (i.e., FC > 2 represents increased expression).** | | | | | |
| **Ensembl ID** | **Gene Symbol** | **Log2FoldChange** | **p-value** | **FDR** | **Signif** |
| ENSG00000147647.13 | DPYS | -3.9E+00 | 5.8E-06 | 4.2E-05 | \* |
| ENSG00000198838.14 | RYR3 | -3.5E+00 | 9.2E-04 | 3.5E-03 | \* |
| ENSG00000162782.16 | TDRD5 | -3.4E+00 | 1.1E-03 | 4.2E-03 | \* |
| ENSG00000152779.14 | SLC16A12 | -3.4E+00 | 8.7E-09 | 1.2E-07 | \* |
| ENSG00000039537.14 | C6 | -3.4E+00 | 4.6E-03 | 1.4E-02 | \* |
| ENSG00000280780.2 | JAKMIP2-AS1 | -3.3E+00 | 8.1E-12 | 2.6E-10 | \* |
| ENSG00000244067.3 | GSTA2 | -3.3E+00 | 1.7E-13 | 8.3E-12 | \* |
| ENSG00000260951.2 | AC005100.1 | -3.2E+00 | 1.5E-05 | 9.4E-05 | \* |
| ENSG00000277893.2 | SRD5A2 | -3.1E+00 | 1.4E-08 | 1.9E-07 | \* |
| ENSG00000268566.6 | AC100781.1 | -3.1E+00 | 6.5E-04 | 2.6E-03 | \* |
| ENSG00000178473.7 | UCN3 | -3.1E+00 | 1.1E-08 | 1.6E-07 | \* |
| ENSG00000144834.14 | TAGLN3 | -3.0E+00 | 1.1E-10 | 2.6E-09 | \* |
| ENSG00000174898.16 | CATSPERD | -3.0E+00 | 1.2E-15 | 1.1E-13 | \* |
| ENSG00000160472.5 | TMEM190 | -3.0E+00 | 1.4E-06 | 1.1E-05 | \* |
| ENSG00000170703.15 | TTLL6 | -3.0E+00 | 8.8E-13 | 3.6E-11 | \* |
| ENSG00000118407.15 | FILIP1 | -3.0E+00 | 1.2E-08 | 1.6E-07 | \* |
| ENSG00000237423.3 | LINC01522 | -3.0E+00 | 7.0E-09 | 1.0E-07 | \* |
| ENSG00000019505.8 | SYT13 | -3.0E+00 | 1.4E-03 | 4.9E-03 | \* |
| ENSG00000175664.10 | TEX26 | -2.9E+00 | 1.0E-11 | 3.1E-10 | \* |
| ENSG00000282904.2 | AC107398.4 | -2.9E+00 | 1.8E-03 | 6.4E-03 | \* |
| ENSG00000197748.13 | CFAP43 | -2.9E+00 | 4.5E-11 | 1.2E-09 | \* |
| ENSG00000150628.7 | SPATA4 | -2.9E+00 | 4.0E-11 | 1.1E-09 | \* |
| ENSG00000178645.13 | C10orf53 | -2.9E+00 | 2.5E-07 | 2.6E-06 | \* |
| ENSG00000232415.1 | ELN-AS1 | -2.9E+00 | 2.8E-12 | 1.0E-10 | \* |
| ENSG00000270332.2 | SMC2-AS1 | -2.9E+00 | 2.3E-11 | 6.5E-10 | \* |
| ENSG00000124490.14 | CRISP2 | -2.9E+00 | 8.6E-07 | 7.6E-06 | \* |
| ENSG00000007174.18 | DNAH9 | -2.9E+00 | 1.7E-11 | 5.1E-10 | \* |
| ENSG00000168878.19 | SFTPB | -2.9E+00 | 4.5E-06 | 3.3E-05 | \* |
| ENSG00000162598.13 | C1orf87 | -2.9E+00 | 1.8E-10 | 4.0E-09 | \* |
| ENSG00000285930.1 | AC015813.8 | -2.9E+00 | 3.2E-11 | 8.5E-10 | \* |
| ENSG00000239389.8 | PCDHA13 | -2.9E+00 | 3.3E-05 | 1.9E-04 | \* |
| ENSG00000226594.1 | AL356421.2 | -2.9E+00 | 2.8E-07 | 2.8E-06 | \* |
| ENSG00000228695.10 | CES1P1 | -2.9E+00 | 3.1E-03 | 1.0E-02 | \* |
| ENSG00000172955.17 | ADH6 | -2.9E+00 | 2.8E-16 | 3.0E-14 | \* |
| ENSG00000246250.3 | AC087521.2 | -2.9E+00 | 4.2E-13 | 1.8E-11 | \* |
| ENSG00000282939.1 | TRBV7-2 | -2.8E+00 | 1.5E-03 | 5.2E-03 | \* |
| ENSG00000171815.6 | PCDHB1 | -2.8E+00 | 4.8E-14 | 2.6E-12 | \* |
| ENSG00000179902.13 | C1orf194 | -2.8E+00 | 2.7E-10 | 5.6E-09 | \* |
| ENSG00000248801.7 | C8orf34-AS1 | -2.8E+00 | 7.9E-04 | 3.1E-03 | \* |
| ENSG00000206531.10 | CD200R1L | -2.8E+00 | 3.0E-03 | 9.7E-03 | \* |
| ENSG00000287189.1 | AL121956.6 | -2.8E+00 | 5.4E-18 | 9.1E-16 | \* |
| ENSG00000117501.14 | MROH9 | -2.8E+00 | 9.1E-13 | 3.7E-11 | \* |
| ENSG00000231720.1 | AL353747.3 | -2.8E+00 | 4.6E-05 | 2.6E-04 | \* |
| ENSG00000260057.6 | LINC01571 | -2.8E+00 | 4.3E-09 | 6.6E-08 | \* |
| ENSG00000231980.1 | AC011899.1 | -2.8E+00 | 3.5E-05 | 2.0E-04 | \* |
| ENSG00000275678.1 | AL133320.2 | -2.8E+00 | 5.0E-05 | 2.8E-04 | \* |
| ENSG00000259508.1 | AL132801.1 | -2.8E+00 | 1.1E-11 | 3.3E-10 | \* |
| ENSG00000182013.18 | PNMA8A | -2.8E+00 | 2.1E-06 | 1.7E-05 | \* |
| ENSG00000230062.6 | ANKRD66 | -2.8E+00 | 1.3E-11 | 4.0E-10 | \* |
| ENSG00000128536.16 | CDHR3 | -2.7E+00 | 1.1E-12 | 4.5E-11 | \* |
| ENSG00000286449.1 | AC016590.4 | -2.7E+00 | 2.9E-05 | 1.7E-04 | \* |
| ENSG00000232192.1 | KIF26B-AS1 | -2.7E+00 | 5.4E-06 | 3.9E-05 | \* |
| ENSG00000187492.9 | CDHR4 | -2.7E+00 | 3.5E-08 | 4.3E-07 | \* |
| ENSG00000157423.18 | HYDIN | -2.7E+00 | 8.7E-09 | 1.2E-07 | \* |
| ENSG00000132464.13 | ENAM | -2.7E+00 | 1.5E-04 | 7.4E-04 | \* |
| ENSG00000188817.8 | SNTN | -2.7E+00 | 1.1E-10 | 2.6E-09 | \* |
| ENSG00000166959.8 | MS4A8 | -2.7E+00 | 1.8E-07 | 1.9E-06 | \* |
| ENSG00000118492.18 | ADGB | -2.7E+00 | 9.1E-10 | 1.7E-08 | \* |
| ENSG00000100012.12 | SEC14L3 | -2.7E+00 | 2.0E-12 | 7.5E-11 | \* |
| ENSG00000173627.8 | APOBEC4 | -2.7E+00 | 6.1E-12 | 2.0E-10 | \* |
| ENSG00000171533.12 | MAP6 | -2.7E+00 | 6.0E-15 | 4.4E-13 | \* |
| ENSG00000286052.1 | AL109955.1 | -2.7E+00 | 4.5E-05 | 2.5E-04 | \* |
| ENSG00000119147.10 | ECRG4 | -2.7E+00 | 1.5E-08 | 2.1E-07 | \* |
| ENSG00000183644.13 | HOATZ | -2.7E+00 | 2.4E-08 | 3.1E-07 | \* |
| ENSG00000184471.8 | C1QTNF8 | -2.7E+00 | 7.1E-06 | 5.0E-05 | \* |
| ENSG00000137473.19 | TTC29 | -2.7E+00 | 1.7E-08 | 2.2E-07 | \* |
| ENSG00000122735.16 | DNAI1 | -2.7E+00 | 1.3E-08 | 1.8E-07 | \* |
| ENSG00000157330.10 | C1orf158 | -2.7E+00 | 2.1E-10 | 4.6E-09 | \* |
| ENSG00000137960.6 | GIPC2 | -2.7E+00 | 2.3E-11 | 6.5E-10 | \* |
| ENSG00000178965.14 | ERICH3 | -2.7E+00 | 3.1E-10 | 6.4E-09 | \* |
| ENSG00000284612.1 | AL591543.1 | -2.7E+00 | 1.8E-03 | 6.3E-03 | \* |
| ENSG00000112539.15 | C6orf118 | -2.7E+00 | 2.7E-10 | 5.6E-09 | \* |
| ENSG00000187726.9 | DNAJB13 | -2.7E+00 | 1.8E-10 | 4.0E-09 | \* |
| ENSG00000068615.19 | REEP1 | -2.6E+00 | 1.7E-09 | 3.0E-08 | \* |
| ENSG00000287003.1 | AC008674.1 | -2.6E+00 | 1.2E-09 | 2.1E-08 | \* |
| ENSG00000146221.10 | TCTE1 | -2.6E+00 | 4.1E-14 | 2.3E-12 | \* |
| ENSG00000115423.19 | DNAH6 | -2.6E+00 | 2.0E-13 | 9.8E-12 | \* |
| ENSG00000186231.17 | KLHL32 | -2.6E+00 | 1.2E-12 | 4.6E-11 | \* |
| ENSG00000163060.8 | TEKT4 | -2.6E+00 | 1.7E-11 | 5.0E-10 | \* |
| ENSG00000143595.13 | AQP10 | -2.6E+00 | 2.6E-10 | 5.5E-09 | \* |
| ENSG00000173988.13 | LRRC63 | -2.6E+00 | 2.6E-06 | 2.0E-05 | \* |
| ENSG00000132554.20 | RGS22 | -2.6E+00 | 1.2E-10 | 2.7E-09 | \* |
| ENSG00000102230.14 | PCYT1B | -2.6E+00 | 2.0E-11 | 5.6E-10 | \* |
| ENSG00000156076.10 | WIF1 | -2.6E+00 | 2.3E-04 | 1.1E-03 | \* |
| ENSG00000276842.1 | AC023510.2 | -2.6E+00 | 6.2E-12 | 2.0E-10 | \* |
| ENSG00000203734.13 | ECT2L | -2.6E+00 | 9.3E-13 | 3.8E-11 | \* |
| ENSG00000109182.12 | CWH43 | -2.6E+00 | 1.1E-04 | 5.5E-04 | \* |
| ENSG00000160188.10 | RSPH1 | -2.6E+00 | 2.5E-14 | 1.5E-12 | \* |
| ENSG00000230710.2 | LINC00332 | -2.6E+00 | 3.6E-04 | 1.5E-03 | \* |
| ENSG00000165309.14 | ARMC3 | -2.6E+00 | 3.5E-08 | 4.3E-07 | \* |
| ENSG00000228723.6 | SRGAP3-AS2 | -2.6E+00 | 3.4E-10 | 6.8E-09 | \* |
| ENSG00000163491.16 | NEK10 | -2.6E+00 | 3.1E-15 | 2.5E-13 | \* |
| ENSG00000179813.7 | FAM216B | -2.6E+00 | 4.5E-09 | 6.9E-08 | \* |
| ENSG00000105278.12 | ZFR2 | -2.6E+00 | 3.5E-05 | 2.0E-04 | \* |
| ENSG00000105479.16 | CCDC114 | -2.6E+00 | 1.3E-07 | 1.4E-06 | \* |
| ENSG00000162643.13 | DNAI3 | -2.6E+00 | 6.1E-09 | 9.0E-08 | \* |
| ENSG00000065534.19 | MYLK | -2.6E+00 | 3.3E-12 | 1.1E-10 | \* |
| ENSG00000230873.9 | STMND1 | -2.5E+00 | 1.2E-10 | 2.7E-09 | \* |
| ENSG00000279965.1 | AL390755.2 | -2.5E+00 | 9.5E-11 | 2.2E-09 | \* |
| ENSG00000133115.12 | STOML3 | -2.5E+00 | 2.8E-08 | 3.6E-07 | \* |
| ENSG00000173947.14 | PIFO | -2.5E+00 | 1.7E-11 | 4.9E-10 | \* |
| ENSG00000275944.1 | AC244100.3 | -2.5E+00 | 1.4E-09 | 2.4E-08 | \* |
| ENSG00000256209.2 | AC073578.1 | -2.5E+00 | 2.0E-02 | 4.8E-02 | \* |
| ENSG00000122420.10 | PTGFR | -2.5E+00 | 6.4E-13 | 2.7E-11 | \* |
| ENSG00000170324.21 | FRMPD2 | -2.5E+00 | 1.1E-08 | 1.5E-07 | \* |
| ENSG00000204815.10 | TTC25 | -2.5E+00 | 7.8E-11 | 1.9E-09 | \* |
| ENSG00000171962.18 | DRC3 | -2.5E+00 | 8.7E-13 | 3.6E-11 | \* |
| ENSG00000253474.2 | AC044893.1 | -2.5E+00 | 1.3E-05 | 8.3E-05 | \* |
| ENSG00000164185.6 | ZNF474 | -2.5E+00 | 2.3E-11 | 6.3E-10 | \* |
| ENSG00000164972.13 | C9orf24 | -2.5E+00 | 3.4E-08 | 4.3E-07 | \* |
| ENSG00000186952.15 | TMEM232 | -2.5E+00 | 4.2E-15 | 3.3E-13 | \* |
| ENSG00000188452.14 | CERKL | -2.5E+00 | 7.3E-09 | 1.1E-07 | \* |
| ENSG00000170482.17 | SLC23A1 | -2.5E+00 | 1.8E-13 | 8.5E-12 | \* |
| ENSG00000110900.16 | TSPAN11 | -2.5E+00 | 3.9E-07 | 3.7E-06 | \* |
| ENSG00000123977.10 | DAW1 | -2.5E+00 | 1.3E-06 | 1.1E-05 | \* |
| ENSG00000149201.10 | CCDC81 | -2.5E+00 | 1.8E-13 | 8.9E-12 | \* |
| ENSG00000184385.2 | UMODL1-AS1 | -2.5E+00 | 9.2E-15 | 6.3E-13 | \* |
| ENSG00000260220.7 | CCDC187 | -2.5E+00 | 2.1E-11 | 6.0E-10 | \* |
| ENSG00000165084.16 | C8orf34 | -2.5E+00 | 1.5E-04 | 7.3E-04 | \* |
| ENSG00000205240.4 | OR7E36P | -2.5E+00 | 5.2E-11 | 1.3E-09 | \* |
| ENSG00000247311.3 | AC010255.1 | -2.5E+00 | 1.2E-10 | 2.7E-09 | \* |
| ENSG00000163576.18 | EFHB | -2.5E+00 | 8.1E-13 | 3.3E-11 | \* |
| ENSG00000286277.1 | AL009177.1 | -2.5E+00 | 7.6E-10 | 1.4E-08 | \* |
| ENSG00000215160.3 | OR7E122P | -2.5E+00 | 2.3E-06 | 1.8E-05 | \* |
| ENSG00000236833.1 | AC024560.1 | -2.5E+00 | 4.5E-03 | 1.4E-02 | \* |
| ENSG00000187151.7 | ANGPTL5 | -2.5E+00 | 1.1E-07 | 1.2E-06 | \* |
| ENSG00000185055.11 | EFCAB10 | -2.5E+00 | 1.9E-12 | 7.3E-11 | \* |
| ENSG00000162814.11 | SPATA17 | -2.4E+00 | 2.6E-12 | 9.5E-11 | \* |
| ENSG00000230368.2 | FAM41C | -2.4E+00 | 8.7E-03 | 2.4E-02 | \* |
| ENSG00000171798.18 | KNDC1 | -2.4E+00 | 1.2E-11 | 3.7E-10 | \* |
| ENSG00000183346.9 | CABCOCO1 | -2.4E+00 | 1.1E-10 | 2.6E-09 | \* |
| ENSG00000157150.5 | TIMP4 | -2.4E+00 | 2.9E-15 | 2.4E-13 | \* |
| ENSG00000234911.2 | TEX21P | -2.4E+00 | 8.9E-15 | 6.2E-13 | \* |
| ENSG00000158445.10 | KCNB1 | -2.4E+00 | 5.8E-09 | 8.7E-08 | \* |
| ENSG00000181378.14 | CFAP65 | -2.4E+00 | 1.3E-10 | 3.0E-09 | \* |
| ENSG00000263450.1 | AC090371.1 | -2.4E+00 | 5.3E-08 | 6.2E-07 | \* |
| ENSG00000222046.2 | DCDC2B | -2.4E+00 | 7.0E-13 | 2.9E-11 | \* |
| ENSG00000224049.1 | AC108681.1 | -2.4E+00 | 3.0E-04 | 1.3E-03 | \* |
| ENSG00000154479.13 | CCDC173 | -2.4E+00 | 2.6E-17 | 3.7E-15 | \* |
| ENSG00000249241.1 | LINC02265 | -2.4E+00 | 1.2E-10 | 2.8E-09 | \* |
| ENSG00000243910.8 | TUBA4B | -2.4E+00 | 8.2E-09 | 1.2E-07 | \* |
| ENSG00000130176.8 | CNN1 | -2.4E+00 | 4.4E-06 | 3.2E-05 | \* |
| ENSG00000151892.15 | GFRA1 | -2.4E+00 | 1.7E-04 | 8.1E-04 | \* |
| ENSG00000080572.13 | DNAAF6 | -2.4E+00 | 1.3E-11 | 3.8E-10 | \* |
| ENSG00000188523.9 | CFAP77 | -2.4E+00 | 4.9E-09 | 7.5E-08 | \* |
| ENSG00000186329.9 | TMEM212 | -2.4E+00 | 3.2E-11 | 8.5E-10 | \* |
| ENSG00000166596.15 | CFAP52 | -2.4E+00 | 2.4E-10 | 5.0E-09 | \* |
| ENSG00000152611.12 | CAPSL | -2.4E+00 | 4.0E-11 | 1.1E-09 | \* |
| ENSG00000196169.15 | KIF19 | -2.4E+00 | 4.2E-11 | 1.1E-09 | \* |
| ENSG00000089101.19 | CFAP61 | -2.4E+00 | 2.1E-13 | 9.8E-12 | \* |
| ENSG00000104237.11 | RP1 | -2.4E+00 | 3.8E-16 | 4.0E-14 | \* |
| ENSG00000204283.4 | LINC01973 | -2.4E+00 | 3.5E-04 | 1.5E-03 | \* |
| ENSG00000197653.16 | DNAH10 | -2.4E+00 | 6.9E-17 | 8.5E-15 | \* |
| ENSG00000111834.13 | RSPH4A | -2.4E+00 | 5.1E-11 | 1.3E-09 | \* |
| ENSG00000155761.14 | SPAG17 | -2.4E+00 | 1.2E-12 | 4.8E-11 | \* |
| ENSG00000153789.13 | CIBAR2 | -2.4E+00 | 4.8E-11 | 1.2E-09 | \* |
| ENSG00000144031.12 | ANKRD53 | -2.4E+00 | 7.0E-12 | 2.3E-10 | \* |
| ENSG00000160401.15 | CFAP157 | -2.4E+00 | 4.1E-07 | 4.0E-06 | \* |
| ENSG00000236495.2 | AL158168.1 | -2.4E+00 | 3.9E-04 | 1.7E-03 | \* |
| ENSG00000231738.11 | TSPAN19 | -2.4E+00 | 1.1E-07 | 1.2E-06 | \* |
| ENSG00000181085.15 | MAPK15 | -2.4E+00 | 3.5E-07 | 3.4E-06 | \* |
| ENSG00000215475.5 | SIAH3 | -2.4E+00 | 1.7E-09 | 2.9E-08 | \* |
| ENSG00000111218.12 | PRMT8 | -2.4E+00 | 6.8E-07 | 6.1E-06 | \* |
| ENSG00000187513.9 | GJA4 | -2.4E+00 | 5.0E-04 | 2.1E-03 | \* |
| ENSG00000174156.15 | GSTA3 | -2.4E+00 | 4.8E-06 | 3.5E-05 | \* |
| ENSG00000260908.1 | AC009093.3 | -2.4E+00 | 3.9E-09 | 6.0E-08 | \* |
| ENSG00000188869.13 | TMC3 | -2.3E+00 | 1.2E-04 | 6.0E-04 | \* |
| ENSG00000278338.4 | VWA8-AS1 | -2.3E+00 | 2.6E-12 | 9.2E-11 | \* |
| ENSG00000034239.11 | EFCAB1 | -2.3E+00 | 5.3E-07 | 4.9E-06 | \* |
| ENSG00000168658.19 | VWA3B | -2.3E+00 | 2.8E-12 | 1.0E-10 | \* |
| ENSG00000124237.6 | C20orf85 | -2.3E+00 | 5.2E-07 | 4.9E-06 | \* |
| ENSG00000125046.15 | SSUH2 | -2.3E+00 | 4.6E-04 | 1.9E-03 | \* |
| ENSG00000149575.9 | SCN2B | -2.3E+00 | 6.0E-05 | 3.3E-04 | \* |
| ENSG00000155026.16 | RSPH10B | -2.3E+00 | 1.7E-10 | 3.7E-09 | \* |
| ENSG00000148408.13 | CACNA1B | -2.3E+00 | 1.1E-04 | 5.4E-04 | \* |
| ENSG00000197057.10 | DTHD1 | -2.3E+00 | 4.7E-15 | 3.6E-13 | \* |
| ENSG00000255974.8 | CYP2A6 | -2.3E+00 | 4.5E-04 | 1.9E-03 | \* |
| ENSG00000137707.13 | BTG4 | -2.3E+00 | 7.6E-09 | 1.1E-07 | \* |
| ENSG00000177994.16 | C2orf73 | -2.3E+00 | 5.6E-06 | 4.0E-05 | \* |
| ENSG00000169515.8 | CCDC8 | -2.3E+00 | 3.9E-08 | 4.7E-07 | \* |
| ENSG00000145423.5 | SFRP2 | -2.3E+00 | 6.8E-05 | 3.6E-04 | \* |
| ENSG00000169126.16 | ARMC4 | -2.3E+00 | 1.1E-10 | 2.6E-09 | \* |
| ENSG00000135333.15 | EPHA7 | -2.3E+00 | 5.3E-04 | 2.2E-03 | \* |
| ENSG00000133640.20 | LRRIQ1 | -2.3E+00 | 9.5E-13 | 3.8E-11 | \* |
| ENSG00000146038.12 | DCDC2 | -2.3E+00 | 2.7E-09 | 4.3E-08 | \* |
| ENSG00000237674.1 | GSTA7P | -2.3E+00 | 2.4E-03 | 8.2E-03 | \* |
| ENSG00000260156.1 | AC020763.1 | -2.3E+00 | 2.0E-07 | 2.0E-06 | \* |
| ENSG00000101448.14 | EPPIN | -2.3E+00 | 7.8E-10 | 1.4E-08 | \* |
| ENSG00000102466.16 | FGF14 | -2.3E+00 | 7.2E-04 | 2.8E-03 | \* |
| ENSG00000232862.5 | AL138787.1 | -2.3E+00 | 2.1E-09 | 3.6E-08 | \* |
| ENSG00000204950.4 | LRRC10B | -2.3E+00 | 4.8E-10 | 9.3E-09 | \* |
| ENSG00000108852.15 | MPP2 | -2.3E+00 | 6.5E-08 | 7.5E-07 | \* |
| ENSG00000215217.7 | C5orf49 | -2.3E+00 | 2.5E-07 | 2.5E-06 | \* |
| ENSG00000226026.6 | AC092802.1 | -2.3E+00 | 9.3E-16 | 8.7E-14 | \* |
| ENSG00000198003.12 | CCDC151 | -2.3E+00 | 6.5E-10 | 1.2E-08 | \* |
| ENSG00000260266.1 | PPIAP46 | -2.3E+00 | 1.3E-04 | 6.3E-04 | \* |
| ENSG00000215187.12 | FAM166B | -2.3E+00 | 2.5E-11 | 7.0E-10 | \* |
| ENSG00000259462.3 | CPEB1-AS1 | -2.3E+00 | 2.1E-06 | 1.7E-05 | \* |
| ENSG00000286339.1 | Z99496.1 | -2.3E+00 | 1.1E-10 | 2.7E-09 | \* |
| ENSG00000140481.15 | CCDC33 | -2.3E+00 | 2.6E-15 | 2.1E-13 | \* |
| ENSG00000186973.11 | FAM183A | -2.3E+00 | 3.8E-14 | 2.1E-12 | \* |
| ENSG00000133665.13 | DYDC2 | -2.3E+00 | 9.3E-08 | 1.0E-06 | \* |
| ENSG00000258752.2 | AL357093.2 | -2.3E+00 | 1.4E-10 | 3.1E-09 | \* |
| ENSG00000251576.1 | LINC01267 | -2.3E+00 | 2.9E-12 | 1.0E-10 | \* |
| ENSG00000150594.7 | ADRA2A | -2.3E+00 | 5.9E-04 | 2.4E-03 | \* |
| ENSG00000237336.1 | AL445489.1 | -2.3E+00 | 1.7E-10 | 3.7E-09 | \* |
| ENSG00000249621.1 | AC113349.1 | -2.3E+00 | 1.3E-09 | 2.3E-08 | \* |
| ENSG00000129654.8 | FOXJ1 | -2.3E+00 | 2.0E-11 | 5.8E-10 | \* |
| ENSG00000239831.1 | RNF7P1 | -2.3E+00 | 1.0E-04 | 5.3E-04 | \* |
| ENSG00000152763.17 | DNAI4 | -2.3E+00 | 9.2E-14 | 4.7E-12 | \* |
| ENSG00000176009.3 | ASCL3 | -2.3E+00 | 4.8E-05 | 2.7E-04 | \* |
| ENSG00000176381.6 | PRR18 | -2.3E+00 | 2.0E-11 | 5.8E-10 | \* |
| ENSG00000166960.16 | CCDC178 | -2.3E+00 | 3.9E-05 | 2.2E-04 | \* |
| ENSG00000187905.10 | LRRC74B | -2.3E+00 | 2.1E-04 | 9.9E-04 | \* |
| ENSG00000165164.14 | CFAP47 | -2.3E+00 | 1.2E-10 | 2.7E-09 | \* |
| ENSG00000237292.1 | LINC01732 | -2.3E+00 | 3.3E-08 | 4.1E-07 | \* |
| ENSG00000274698.1 | AC099521.1 | -2.3E+00 | 3.4E-08 | 4.2E-07 | \* |
| ENSG00000173838.12 | MARCHF10 | -2.3E+00 | 1.1E-15 | 9.9E-14 | \* |
| ENSG00000186471.13 | AKAP14 | -2.3E+00 | 5.4E-08 | 6.3E-07 | \* |
| ENSG00000159713.11 | TPPP3 | -2.3E+00 | 2.3E-15 | 1.9E-13 | \* |
| ENSG00000243836.5 | WDR86-AS1 | -2.3E+00 | 1.3E-10 | 2.9E-09 | \* |
| ENSG00000139537.11 | CCDC65 | -2.3E+00 | 4.8E-10 | 9.3E-09 | \* |
| ENSG00000243710.8 | CFAP57 | -2.3E+00 | 2.1E-12 | 7.6E-11 | \* |
| ENSG00000181322.15 | NME9 | -2.2E+00 | 5.2E-11 | 1.3E-09 | \* |
| ENSG00000178233.18 | TMEM151B | -2.2E+00 | 1.1E-03 | 4.2E-03 | \* |
| ENSG00000186976.15 | EFCAB6 | -2.2E+00 | 1.0E-16 | 1.2E-14 | \* |
| ENSG00000166246.14 | C16orf71 | -2.2E+00 | 3.5E-16 | 3.7E-14 | \* |
| ENSG00000287600.1 | AC122719.3 | -2.2E+00 | 1.3E-10 | 3.0E-09 | \* |
| ENSG00000231533.2 | AL391840.1 | -2.2E+00 | 3.6E-08 | 4.4E-07 | \* |
| ENSG00000170231.16 | FABP6 | -2.2E+00 | 1.6E-08 | 2.1E-07 | \* |
| ENSG00000153822.14 | KCNJ16 | -2.2E+00 | 3.3E-03 | 1.1E-02 | \* |
| ENSG00000178796.13 | RIIAD1 | -2.2E+00 | 5.1E-10 | 9.8E-09 | \* |
| ENSG00000138400.13 | MDH1B | -2.2E+00 | 1.3E-12 | 5.0E-11 | \* |
| ENSG00000142609.18 | CFAP74 | -2.2E+00 | 9.3E-09 | 1.3E-07 | \* |
| ENSG00000174844.14 | DNAH12 | -2.2E+00 | 1.1E-17 | 1.7E-15 | \* |
| ENSG00000183631.5 | PRR32 | -2.2E+00 | 1.7E-02 | 4.2E-02 | \* |
| ENSG00000006611.17 | USH1C | -2.2E+00 | 1.4E-06 | 1.2E-05 | \* |
| ENSG00000180383.3 | DEFB124 | -2.2E+00 | 4.6E-05 | 2.6E-04 | \* |
| ENSG00000144583.5 | MARCHF4 | -2.2E+00 | 4.0E-05 | 2.3E-04 | \* |
| ENSG00000116039.13 | ATP6V1B1 | -2.2E+00 | 2.9E-07 | 2.9E-06 | \* |
| ENSG00000279725.1 | AL391005.1 | -2.2E+00 | 3.0E-05 | 1.8E-04 | \* |
| ENSG00000226690.8 | AC013470.2 | -2.2E+00 | 4.4E-06 | 3.2E-05 | \* |
| ENSG00000165621.8 | OXGR1 | -2.2E+00 | 1.5E-09 | 2.6E-08 | \* |
| ENSG00000156206.14 | CFAP161 | -2.2E+00 | 4.0E-13 | 1.7E-11 | \* |
| ENSG00000203985.11 | LDLRAD1 | -2.2E+00 | 4.3E-16 | 4.4E-14 | \* |
| ENSG00000167646.14 | DNAAF3 | -2.2E+00 | 3.5E-11 | 9.4E-10 | \* |
| ENSG00000185681.13 | MORN5 | -2.2E+00 | 1.3E-12 | 4.9E-11 | \* |
| ENSG00000069122.19 | ADGRF5 | -2.2E+00 | 2.4E-07 | 2.4E-06 | \* |
| ENSG00000224383.8 | PRR29 | -2.2E+00 | 1.0E-10 | 2.5E-09 | \* |
| ENSG00000162004.18 | CCDC78 | -2.2E+00 | 3.8E-13 | 1.7E-11 | \* |
| ENSG00000259362.2 | AC046168.1 | -2.2E+00 | 1.7E-03 | 6.0E-03 | \* |
| ENSG00000152936.10 | LMNTD1 | -2.2E+00 | 6.0E-13 | 2.6E-11 | \* |
| ENSG00000214575.9 | CPEB1 | -2.2E+00 | 3.7E-06 | 2.8E-05 | \* |
| ENSG00000141665.13 | FBXO15 | -2.2E+00 | 3.8E-12 | 1.3E-10 | \* |
| ENSG00000155816.21 | FMN2 | -2.2E+00 | 1.5E-03 | 5.4E-03 | \* |
| ENSG00000269091.6 | AC010624.3 | -2.2E+00 | 3.8E-07 | 3.7E-06 | \* |
| ENSG00000188396.4 | TCTEX1D4 | -2.2E+00 | 3.0E-10 | 6.2E-09 | \* |
| ENSG00000286028.1 | AP002008.4 | -2.2E+00 | 7.2E-07 | 6.5E-06 | \* |
| ENSG00000242908.6 | AADACL2-AS1 | -2.2E+00 | 1.8E-10 | 4.0E-09 | \* |
| ENSG00000162543.6 | UBXN10 | -2.2E+00 | 1.8E-12 | 6.7E-11 | \* |
| ENSG00000179071.5 | CCDC89 | -2.2E+00 | 4.2E-09 | 6.4E-08 | \* |
| ENSG00000145002.12 | FAM86B2 | -2.2E+00 | 3.0E-04 | 1.3E-03 | \* |
| ENSG00000231057.4 | AC096637.2 | -2.2E+00 | 2.5E-09 | 4.1E-08 | \* |
| ENSG00000182329.14 | KIAA2012 | -2.2E+00 | 1.9E-09 | 3.2E-08 | \* |
| ENSG00000183833.16 | CFAP91 | -2.2E+00 | 2.0E-08 | 2.6E-07 | \* |
| ENSG00000257108.2 | NHLRC4 | -2.2E+00 | 1.5E-11 | 4.6E-10 | \* |
| ENSG00000077327.16 | SPAG6 | -2.2E+00 | 1.2E-06 | 9.9E-06 | \* |
| ENSG00000231106.2 | LINC01436 | -2.2E+00 | 1.7E-03 | 5.9E-03 | \* |
| ENSG00000169402.15 | RSPH10B2 | -2.2E+00 | 6.3E-07 | 5.7E-06 | \* |
| ENSG00000112530.11 | PACRG | -2.2E+00 | 9.8E-09 | 1.4E-07 | \* |
| ENSG00000137691.13 | CFAP300 | -2.2E+00 | 2.4E-11 | 6.5E-10 | \* |
| ENSG00000114455.13 | HHLA2 | -2.2E+00 | 4.7E-08 | 5.6E-07 | \* |
| ENSG00000153347.10 | FAM81B | -2.2E+00 | 2.0E-10 | 4.3E-09 | \* |
| ENSG00000171695.10 | LKAAEAR1 | -2.2E+00 | 9.3E-04 | 3.6E-03 | \* |
| ENSG00000188316.15 | ENO4 | -2.2E+00 | 3.8E-09 | 5.9E-08 | \* |
| ENSG00000165182.12 | CXorf58 | -2.2E+00 | 5.0E-11 | 1.3E-09 | \* |
| ENSG00000138587.6 | MNS1 | -2.2E+00 | 7.5E-11 | 1.8E-09 | \* |
| ENSG00000116032.5 | GRIN3B | -2.2E+00 | 1.1E-10 | 2.6E-09 | \* |
| ENSG00000270765.6 | GAS2L2 | -2.2E+00 | 2.3E-11 | 6.4E-10 | \* |
| ENSG00000132259.13 | CNGA4 | -2.2E+00 | 2.5E-11 | 6.8E-10 | \* |
| ENSG00000188039.16 | NWD1 | -2.2E+00 | 1.5E-26 | 1.1E-23 | \* |
| ENSG00000176601.13 | MAP3K19 | -2.2E+00 | 9.6E-07 | 8.3E-06 | \* |
| ENSG00000136918.8 | WDR38 | -2.1E+00 | 2.2E-12 | 7.9E-11 | \* |
| ENSG00000255308.1 | CSRP3-AS1 | -2.1E+00 | 1.1E-12 | 4.3E-11 | \* |
| ENSG00000150750.8 | C11orf53 | -2.1E+00 | 2.0E-05 | 1.3E-04 | \* |
| ENSG00000144057.15 | ST6GAL2 | -2.1E+00 | 6.7E-06 | 4.7E-05 | \* |
| ENSG00000287401.1 | AC034139.1 | -2.1E+00 | 4.9E-09 | 7.4E-08 | \* |
| ENSG00000163263.7 | C1orf189 | -2.1E+00 | 5.3E-17 | 6.8E-15 | \* |
| ENSG00000249715.13 | FER1L5 | -2.1E+00 | 1.6E-05 | 1.0E-04 | \* |
| ENSG00000248329.6 | APELA | -2.1E+00 | 1.7E-02 | 4.1E-02 | \* |
| ENSG00000152760.10 | TCTEX1D1 | -2.1E+00 | 1.2E-07 | 1.3E-06 | \* |
| ENSG00000131044.17 | TTLL9 | -2.1E+00 | 9.9E-07 | 8.6E-06 | \* |
| ENSG00000126838.10 | PZP | -2.1E+00 | 2.6E-06 | 2.0E-05 | \* |
| ENSG00000187957.8 | DNER | -2.1E+00 | 2.5E-10 | 5.2E-09 | \* |
| ENSG00000105877.18 | DNAH11 | -2.1E+00 | 9.2E-11 | 2.2E-09 | \* |
| ENSG00000123243.15 | ITIH5 | -2.1E+00 | 3.2E-05 | 1.9E-04 | \* |
| ENSG00000160838.14 | LRRC71 | -2.1E+00 | 5.7E-09 | 8.6E-08 | \* |
| ENSG00000136449.15 | MYCBPAP | -2.1E+00 | 2.8E-11 | 7.6E-10 | \* |
| ENSG00000227620.4 | ALG1L8P | -2.1E+00 | 3.3E-06 | 2.5E-05 | \* |
| ENSG00000125409.13 | TEKT3 | -2.1E+00 | 1.5E-07 | 1.6E-06 | \* |
| ENSG00000205084.11 | TMEM231 | -2.1E+00 | 1.3E-14 | 8.7E-13 | \* |
| ENSG00000146521.10 | LINC01558 | -2.1E+00 | 3.3E-04 | 1.4E-03 | \* |
| ENSG00000088320.4 | REM1 | -2.1E+00 | 5.9E-04 | 2.4E-03 | \* |
| ENSG00000112183.15 | RBM24 | -2.1E+00 | 1.2E-07 | 1.3E-06 | \* |
| ENSG00000130433.8 | CACNG6 | -2.1E+00 | 3.2E-07 | 3.1E-06 | \* |
| ENSG00000162399.9 | BSND | -2.1E+00 | 1.7E-04 | 8.0E-04 | \* |
| ENSG00000118307.20 | CFAP94 | -2.1E+00 | 5.9E-14 | 3.2E-12 | \* |
| ENSG00000188931.4 | CFAP126 | -2.1E+00 | 9.1E-07 | 8.0E-06 | \* |
| ENSG00000229654.1 | LINC02526 | -2.1E+00 | 4.1E-05 | 2.3E-04 | \* |
| ENSG00000171595.14 | DNAI2 | -2.1E+00 | 1.8E-09 | 3.0E-08 | \* |
| ENSG00000174776.11 | WDR49 | -2.1E+00 | 1.5E-13 | 7.6E-12 | \* |
| ENSG00000163879.11 | DNALI1 | -2.1E+00 | 1.8E-08 | 2.4E-07 | \* |
| ENSG00000164627.18 | KIF6 | -2.1E+00 | 8.8E-14 | 4.5E-12 | \* |
| ENSG00000259225.7 | LINC02345 | -2.1E+00 | 2.0E-10 | 4.3E-09 | \* |
| ENSG00000168589.15 | DYNLRB2 | -2.1E+00 | 1.0E-09 | 1.8E-08 | \* |
| ENSG00000185250.16 | PPIL6 | -2.1E+00 | 9.4E-10 | 1.7E-08 | \* |
| ENSG00000277639.2 | AC007906.2 | -2.1E+00 | 6.4E-16 | 6.3E-14 | \* |
| ENSG00000166111.10 | SVOP | -2.1E+00 | 2.1E-04 | 9.7E-04 | \* |
| ENSG00000163075.13 | CFAP221 | -2.1E+00 | 1.4E-08 | 1.9E-07 | \* |
| ENSG00000287435.1 | AC069404.1 | -2.1E+00 | 7.5E-05 | 3.9E-04 | \* |
| ENSG00000140067.6 | FAM181A | -2.1E+00 | 4.9E-09 | 7.5E-08 | \* |
| ENSG00000265752.3 | AC010754.1 | -2.1E+00 | 2.4E-09 | 4.0E-08 | \* |
| ENSG00000137098.14 | SPAG8 | -2.1E+00 | 1.6E-10 | 3.5E-09 | \* |
| ENSG00000175175.6 | PPM1E | -2.1E+00 | 5.8E-08 | 6.8E-07 | \* |
| ENSG00000175267.15 | VWA3A | -2.1E+00 | 5.9E-08 | 6.8E-07 | \* |
| ENSG00000159708.18 | LRRC36 | -2.1E+00 | 8.2E-08 | 9.3E-07 | \* |
| ENSG00000176029.14 | C11orf16 | -2.1E+00 | 7.3E-09 | 1.1E-07 | \* |
| ENSG00000258584.2 | FAM181A-AS1 | -2.1E+00 | 1.0E-08 | 1.4E-07 | \* |
| ENSG00000164746.14 | C7orf57 | -2.1E+00 | 3.8E-11 | 1.0E-09 | \* |
| ENSG00000181123.8 | AC004832.1 | -2.1E+00 | 4.3E-10 | 8.5E-09 | \* |
| ENSG00000118997.14 | DNAH7 | -2.1E+00 | 6.0E-21 | 1.8E-18 | \* |
| ENSG00000126861.5 | OMG | -2.1E+00 | 8.5E-07 | 7.5E-06 | \* |
| ENSG00000129951.19 | PLPPR3 | -2.1E+00 | 5.4E-05 | 3.0E-04 | \* |
| ENSG00000179133.15 | C10orf67 | -2.1E+00 | 1.5E-11 | 4.4E-10 | \* |
| ENSG00000259038.1 | AL121820.2 | -2.1E+00 | 6.8E-11 | 1.7E-09 | \* |
| ENSG00000287117.1 | AC105916.1 | -2.1E+00 | 8.7E-07 | 7.7E-06 | \* |
| ENSG00000204711.9 | C9orf135 | -2.1E+00 | 4.6E-07 | 4.3E-06 | \* |
| ENSG00000265374.1 | LINC01908 | -2.1E+00 | 1.4E-06 | 1.2E-05 | \* |
| ENSG00000242759.8 | LINC00882 | -2.1E+00 | 7.5E-09 | 1.1E-07 | \* |
| ENSG00000163885.12 | CFAP100 | -2.1E+00 | 3.3E-05 | 1.9E-04 | \* |
| ENSG00000255093.2 | AP002008.2 | -2.1E+00 | 8.1E-05 | 4.2E-04 | \* |
| ENSG00000165923.16 | AGBL2 | -2.1E+00 | 6.4E-13 | 2.7E-11 | \* |
| ENSG00000196277.16 | GRM7 | -2.1E+00 | 3.5E-10 | 7.0E-09 | \* |
| ENSG00000142959.5 | BEST4 | -2.1E+00 | 7.4E-05 | 3.9E-04 | \* |
| ENSG00000286330.1 | AL353660.1 | -2.0E+00 | 7.6E-11 | 1.8E-09 | \* |
| ENSG00000249464.6 | LINC01091 | -2.0E+00 | 4.5E-09 | 6.9E-08 | \* |
| ENSG00000172361.6 | CFAP53 | -2.0E+00 | 7.4E-13 | 3.1E-11 | \* |
| ENSG00000186094.17 | AGBL4 | -2.0E+00 | 1.7E-07 | 1.7E-06 | \* |
| ENSG00000258334.1 | AC125611.4 | -2.0E+00 | 3.3E-08 | 4.1E-07 | \* |
| ENSG00000186710.11 | CFAP73 | -2.0E+00 | 5.0E-12 | 1.7E-10 | \* |
| ENSG00000224445.3 | LINC01708 | -2.0E+00 | 6.1E-05 | 3.3E-04 | \* |
| ENSG00000203778.8 | FAM229B | -2.0E+00 | 8.6E-14 | 4.5E-12 | \* |
| ENSG00000206113.11 | CFAP99 | -2.0E+00 | 2.8E-09 | 4.5E-08 | \* |
| ENSG00000173557.15 | FAM166C | -2.0E+00 | 3.0E-09 | 4.8E-08 | \* |
| ENSG00000128408.9 | RIBC2 | -2.0E+00 | 6.3E-10 | 1.2E-08 | \* |
| ENSG00000286830.1 | AC105052.5 | -2.0E+00 | 1.5E-07 | 1.6E-06 | \* |
| ENSG00000196096.3 | SPAG16-DT | -2.0E+00 | 5.7E-06 | 4.1E-05 | \* |
| ENSG00000170959.14 | DCDC1 | -2.0E+00 | 7.4E-09 | 1.1E-07 | \* |
| ENSG00000102904.15 | TSNAXIP1 | -2.0E+00 | 1.0E-09 | 1.8E-08 | \* |
| ENSG00000272514.6 | CFAP206 | -2.0E+00 | 1.5E-06 | 1.2E-05 | \* |
| ENSG00000287312.1 | AC127526.4 | -2.0E+00 | 2.7E-06 | 2.1E-05 | \* |
| ENSG00000259426.6 | AC027237.3 | -2.0E+00 | 7.0E-13 | 3.0E-11 | \* |
| ENSG00000105519.18 | CAPS | -2.0E+00 | 9.3E-19 | 1.9E-16 | \* |
| ENSG00000259087.5 | AL121790.2 | -2.0E+00 | 2.1E-07 | 2.1E-06 | \* |
| ENSG00000137860.12 | SLC28A2 | -2.0E+00 | 9.3E-03 | 2.5E-02 | \* |
| ENSG00000259969.1 | AL049838.1 | -2.0E+00 | 8.1E-12 | 2.6E-10 | \* |
| ENSG00000203963.11 | C1orf141 | -2.0E+00 | 7.1E-03 | 2.0E-02 | \* |
| ENSG00000257057.3 | C11orf97 | -2.0E+00 | 3.6E-05 | 2.1E-04 | \* |
| ENSG00000198848.13 | CES1 | -2.0E+00 | 9.8E-09 | 1.4E-07 | \* |
| ENSG00000239467.6 | AC007405.3 | -2.0E+00 | 5.5E-06 | 3.9E-05 | \* |
| ENSG00000204131.9 | NHSL2 | 2.0E+00 | 1.0E-09 | 1.8E-08 | \* |
| ENSG00000072952.19 | IRAG1 | 2.0E+00 | 6.9E-09 | 1.0E-07 | \* |
| ENSG00000179348.12 | GATA2 | 2.0E+00 | 2.0E-02 | 4.7E-02 | \* |
| ENSG00000140932.10 | CMTM2 | 2.0E+00 | 8.7E-05 | 4.5E-04 | \* |
| ENSG00000128383.13 | APOBEC3A | 2.0E+00 | 3.5E-05 | 2.1E-04 | \* |
| ENSG00000142405.22 | NLRP12 | 2.0E+00 | 1.1E-05 | 7.4E-05 | \* |
| ENSG00000158517.15 | NCF1 | 2.0E+00 | 1.9E-11 | 5.4E-10 | \* |
| ENSG00000225101.6 | OR52K3P | 2.0E+00 | 9.3E-06 | 6.3E-05 | \* |
| ENSG00000286076.1 | AC005050.3 | 2.0E+00 | 1.5E-02 | 3.8E-02 | \* |
| ENSG00000235568.7 | NFAM1 | 2.0E+00 | 8.9E-10 | 1.6E-08 | \* |
| ENSG00000111052.8 | LIN7A | 2.0E+00 | 2.2E-07 | 2.2E-06 | \* |
| ENSG00000261026.1 | AC105046.1 | 2.0E+00 | 4.3E-04 | 1.8E-03 | \* |
| ENSG00000102445.20 | RUBCNL | 2.0E+00 | 2.0E-07 | 2.0E-06 | \* |
| ENSG00000237927.1 | AL078604.2 | 2.0E+00 | 3.2E-03 | 1.0E-02 | \* |
| ENSG00000124731.13 | TREM1 | 2.0E+00 | 3.6E-04 | 1.5E-03 | \* |
| ENSG00000287510.1 | AL137857.1 | 2.0E+00 | 2.0E-04 | 9.2E-04 | \* |
| ENSG00000166523.8 | CLEC4E | 2.0E+00 | 1.3E-04 | 6.3E-04 | \* |
| ENSG00000018280.17 | SLC11A1 | 2.1E+00 | 9.6E-06 | 6.4E-05 | \* |
| ENSG00000123689.6 | G0S2 | 2.1E+00 | 5.1E-05 | 2.8E-04 | \* |
| ENSG00000136695.15 | IL36RN | 2.1E+00 | 2.4E-03 | 7.9E-03 | \* |
| ENSG00000136630.13 | HLX | 2.1E+00 | 1.9E-07 | 2.0E-06 | \* |
| ENSG00000229256.1 | ST13P13 | 2.1E+00 | 6.5E-03 | 1.9E-02 | \* |
| ENSG00000163563.8 | MNDA | 2.1E+00 | 7.7E-06 | 5.3E-05 | \* |
| ENSG00000171051.9 | FPR1 | 2.1E+00 | 3.5E-05 | 2.0E-04 | \* |
| ENSG00000184106.8 | TREML3P | 2.1E+00 | 3.6E-04 | 1.5E-03 | \* |
| ENSG00000158786.5 | PLA2G2F | 2.1E+00 | 1.9E-06 | 1.5E-05 | \* |
| ENSG00000241794.2 | SPRR2A | 2.1E+00 | 9.5E-06 | 6.4E-05 | \* |
| ENSG00000196209.13 | SIRPB2 | 2.1E+00 | 3.0E-08 | 3.7E-07 | \* |
| ENSG00000232528.3 | AL109809.1 | 2.1E+00 | 5.7E-05 | 3.1E-04 | \* |
| ENSG00000189051.5 | RNF222 | 2.1E+00 | 9.7E-06 | 6.5E-05 | \* |
| ENSG00000241163.8 | LINC00877 | 2.1E+00 | 7.3E-06 | 5.1E-05 | \* |
| ENSG00000067646.12 | ZFY | 2.1E+00 | 1.6E-02 | 3.9E-02 | \* |
| ENSG00000186642.16 | PDE2A | 2.1E+00 | 6.5E-07 | 5.9E-06 | \* |
| ENSG00000260943.1 | LINC02555 | 2.1E+00 | 4.7E-03 | 1.4E-02 | \* |
| ENSG00000203785.9 | SPRR2E | 2.1E+00 | 4.0E-04 | 1.7E-03 | \* |
| ENSG00000167755.15 | KLK6 | 2.1E+00 | 5.9E-05 | 3.2E-04 | \* |
| ENSG00000170956.17 | CEACAM3 | 2.1E+00 | 7.3E-09 | 1.1E-07 | \* |
| ENSG00000115008.6 | IL1B | 2.1E+00 | 1.2E-03 | 4.5E-03 | \* |
| ENSG00000113749.7 | HRH2 | 2.1E+00 | 2.3E-08 | 3.0E-07 | \* |
| ENSG00000160883.11 | HK3 | 2.1E+00 | 5.0E-04 | 2.1E-03 | \* |
| ENSG00000154734.16 | ADAMTS1 | 2.1E+00 | 3.4E-04 | 1.5E-03 | \* |
| ENSG00000128917.8 | DLL4 | 2.1E+00 | 2.3E-04 | 1.1E-03 | \* |
| ENSG00000105501.12 | SIGLEC5 | 2.1E+00 | 3.3E-03 | 1.1E-02 | \* |
| ENSG00000112195.9 | TREML2 | 2.1E+00 | 3.6E-05 | 2.1E-04 | \* |
| ENSG00000143631.11 | FLG | 2.2E+00 | 3.8E-03 | 1.2E-02 | \* |
| ENSG00000100368.14 | CSF2RB | 2.2E+00 | 6.1E-07 | 5.6E-06 | \* |
| ENSG00000163823.4 | CCR1 | 2.2E+00 | 3.2E-07 | 3.1E-06 | \* |
| ENSG00000244094.2 | SPRR2F | 2.2E+00 | 8.5E-04 | 3.3E-03 | \* |
| ENSG00000143369.15 | ECM1 | 2.2E+00 | 2.7E-06 | 2.1E-05 | \* |
| ENSG00000151948.12 | GLT1D1 | 2.2E+00 | 1.1E-06 | 9.7E-06 | \* |
| ENSG00000277452.1 | RN7SL473P | 2.2E+00 | 2.7E-05 | 1.6E-04 | \* |
| ENSG00000204577.11 | LILRB3 | 2.2E+00 | 1.0E-07 | 1.1E-06 | \* |
| ENSG00000108342.13 | CSF3 | 2.2E+00 | 1.0E-02 | 2.8E-02 | \* |
| ENSG00000264204.2 | AGAP7P | 2.2E+00 | 3.7E-06 | 2.8E-05 | \* |
| ENSG00000239998.6 | LILRA2 | 2.2E+00 | 2.0E-05 | 1.2E-04 | \* |
| ENSG00000189001.11 | SBSN | 2.2E+00 | 8.3E-05 | 4.3E-04 | \* |
| ENSG00000196136.18 | SERPINA3 | 2.2E+00 | 4.2E-03 | 1.3E-02 | \* |
| ENSG00000101916.12 | TLR8 | 2.2E+00 | 3.0E-06 | 2.3E-05 | \* |
| ENSG00000263934.5 | SNORD3A | 2.2E+00 | 1.1E-03 | 4.1E-03 | \* |
| ENSG00000104974.12 | LILRA1 | 2.3E+00 | 9.9E-08 | 1.1E-06 | \* |
| ENSG00000050730.16 | TNIP3 | 2.3E+00 | 9.7E-04 | 3.7E-03 | \* |
| ENSG00000171777.16 | RASGRP4 | 2.3E+00 | 9.8E-09 | 1.4E-07 | \* |
| ENSG00000105352.10 | CEACAM4 | 2.3E+00 | 9.0E-10 | 1.7E-08 | \* |
| ENSG00000173868.12 | PHOSPHO1 | 2.3E+00 | 1.9E-05 | 1.2E-04 | \* |
| ENSG00000181409.14 | AATK | 2.3E+00 | 4.7E-06 | 3.5E-05 | \* |
| ENSG00000171049.9 | FPR2 | 2.3E+00 | 3.6E-05 | 2.1E-04 | \* |
| ENSG00000135636.14 | DYSF | 2.3E+00 | 9.8E-08 | 1.1E-06 | \* |
| ENSG00000284948.1 | AC107959.4 | 2.3E+00 | 8.3E-05 | 4.3E-04 | \* |
| ENSG00000008516.18 | MMP25 | 2.3E+00 | 2.7E-11 | 7.4E-10 | \* |
| ENSG00000180871.8 | CXCR2 | 2.3E+00 | 6.8E-07 | 6.2E-06 | \* |
| ENSG00000233559.2 | LINC00513 | 2.3E+00 | 4.5E-06 | 3.3E-05 | \* |
| ENSG00000131002.12 | TXLNGY | 2.3E+00 | 3.8E-03 | 1.2E-02 | \* |
| ENSG00000163421.9 | PROK2 | 2.3E+00 | 4.9E-04 | 2.0E-03 | \* |
| ENSG00000184557.4 | SOCS3 | 2.3E+00 | 3.4E-08 | 4.2E-07 | \* |
| ENSG00000162747.12 | FCGR3B | 2.3E+00 | 1.3E-04 | 6.6E-04 | \* |
| ENSG00000136244.12 | IL6 | 2.3E+00 | 1.2E-03 | 4.5E-03 | \* |
| ENSG00000119535.18 | CSF3R | 2.3E+00 | 3.7E-07 | 3.5E-06 | \* |
| ENSG00000287926.1 | AC113208.5 | 2.4E+00 | 1.7E-04 | 8.0E-04 | \* |
| ENSG00000163464.8 | CXCR1 | 2.4E+00 | 5.1E-05 | 2.8E-04 | \* |
| ENSG00000213816.3 | CNN2P4 | 2.4E+00 | 9.2E-04 | 3.5E-03 | \* |
| ENSG00000244482.10 | LILRA6 | 2.4E+00 | 2.5E-06 | 1.9E-05 | \* |
| ENSG00000118520.15 | ARG1 | 2.4E+00 | 6.3E-04 | 2.5E-03 | \* |
| ENSG00000169509.6 | CRCT1 | 2.4E+00 | 1.1E-03 | 4.1E-03 | \* |
| ENSG00000103313.13 | MEFV | 2.4E+00 | 6.4E-07 | 5.8E-06 | \* |
| ENSG00000218809.1 | AL391903.1 | 2.4E+00 | 1.2E-04 | 6.1E-04 | \* |
| ENSG00000174885.13 | NLRP6 | 2.4E+00 | 3.4E-08 | 4.2E-07 | \* |
| ENSG00000124391.5 | IL17C | 2.4E+00 | 1.1E-04 | 5.5E-04 | \* |
| ENSG00000234436.1 | AC245884.2 | 2.4E+00 | 8.6E-04 | 3.3E-03 | \* |
| ENSG00000198019.13 | FCGR1B | 2.4E+00 | 6.8E-06 | 4.8E-05 | \* |
| ENSG00000233013.10 | FAM157B | 2.4E+00 | 1.2E-06 | 1.0E-05 | \* |
| ENSG00000173535.15 | TNFRSF10C | 2.4E+00 | 2.1E-06 | 1.7E-05 | \* |
| ENSG00000126262.5 | FFAR2 | 2.5E+00 | 2.1E-05 | 1.3E-04 | \* |
| ENSG00000225774.2 | SIRPAP1 | 2.5E+00 | 8.5E-03 | 2.3E-02 | \* |
| ENSG00000137757.11 | CASP5 | 2.5E+00 | 2.7E-06 | 2.1E-05 | \* |
| ENSG00000237576.2 | LINC01888 | 2.5E+00 | 8.3E-03 | 2.3E-02 | \* |
| ENSG00000175264.8 | CHST1 | 2.5E+00 | 2.8E-04 | 1.3E-03 | \* |
| ENSG00000251230.6 | MIR3945HG | 2.5E+00 | 1.3E-03 | 4.8E-03 | \* |
| ENSG00000201998.1 | SNORA23 | 2.6E+00 | 8.6E-03 | 2.4E-02 | \* |
| ENSG00000278771.1 | RN7SL3 | 2.6E+00 | 3.1E-04 | 1.4E-03 | \* |
| ENSG00000259600.2 | AC066616.2 | 2.6E+00 | 3.5E-03 | 1.1E-02 | \* |
| ENSG00000085265.11 | FCN1 | 2.6E+00 | 1.6E-04 | 7.9E-04 | \* |
| ENSG00000287937.1 | AC079753.2 | 2.6E+00 | 1.5E-03 | 5.4E-03 | \* |
| ENSG00000249662.6 | LINC02218 | 2.6E+00 | 7.0E-06 | 4.9E-05 | \* |
| ENSG00000250687.6 | NAIPP1 | 2.6E+00 | 1.8E-03 | 6.1E-03 | \* |
| ENSG00000255801.1 | AC092746.1 | 2.6E+00 | 9.1E-04 | 3.5E-03 | \* |
| ENSG00000181333.12 | HEPHL1 | 2.6E+00 | 1.6E-03 | 5.7E-03 | \* |
| ENSG00000268658.5 | LINC00664 | 2.7E+00 | 6.2E-03 | 1.8E-02 | \* |
| ENSG00000249173.6 | LINC01093 | 2.7E+00 | 1.3E-03 | 4.7E-03 | \* |
| ENSG00000200087.1 | SNORA73B | 2.7E+00 | 9.9E-04 | 3.8E-03 | \* |
| ENSG00000255328.1 | AC136475.5 | 2.7E+00 | 8.7E-05 | 4.5E-04 | \* |
| ENSG00000186407.7 | CD300E | 2.7E+00 | 5.4E-03 | 1.6E-02 | \* |
| ENSG00000125538.12 | IL1B | 2.8E+00 | 5.0E-05 | 2.8E-04 | \* |
| ENSG00000255823.5 | MTRNR2L8 | 2.8E+00 | 1.1E-02 | 2.8E-02 | \* |
| ENSG00000197253.13 | TPSB2 | 2.8E+00 | 1.1E-02 | 2.8E-02 | \* |
| ENSG00000274008.1 | Metazoa\_SRP | 2.8E+00 | 6.1E-06 | 4.3E-05 | \* |
| ENSG00000207445.1 | SNORD15B | 2.8E+00 | 6.9E-03 | 2.0E-02 | \* |
| ENSG00000167916.5 | KRT24 | 2.9E+00 | 3.0E-04 | 1.3E-03 | \* |
| ENSG00000264229.1 | RNU4ATAC | 2.9E+00 | 1.1E-02 | 3.0E-02 | \* |
| ENSG00000269495.1 | AC011483.2 | 2.9E+00 | 1.5E-03 | 5.4E-03 | \* |
| ENSG00000202538.1 | RNU4-2 | 2.9E+00 | 1.6E-02 | 3.9E-02 | \* |
| ENSG00000163736.4 | PPBP | 3.1E+00 | 9.3E-07 | 8.1E-06 | \* |
| ENSG00000276168.1 | RN7SL1 | 3.1E+00 | 2.7E-05 | 1.6E-04 | \* |
| ENSG00000201098.1 | RNY1 | 3.3E+00 | 1.0E-03 | 3.8E-03 | \* |
| ENSG00000268240.1 | AC123912.2 | 3.3E+00 | 1.7E-04 | 8.1E-04 | \* |
| ENSG00000212443.1 | SNORA53 | 3.4E+00 | 6.6E-03 | 1.9E-02 | \* |
| ENSG00000183878.15 | UTY | 3.5E+00 | 1.5E-04 | 7.1E-04 | \* |
| ENSG00000201321.1 | RNA5S9 | 3.7E+00 | 3.8E-03 | 1.2E-02 | \* |
| ENSG00000225972.1 | MTND1P23 | 4.2E+00 | 9.3E-04 | 3.6E-03 | \* |
| ENSG00000232177.1 | MTND4P24 | 4.7E+00 | 4.2E-04 | 1.8E-03 | \* |
| ENSG00000114374.13 | USP9Y | 4.8E+00 | 5.8E-03 | 1.7E-02 | \* |
| ENSG00000067048.17 | DDX3Y | 5.7E+00 | 2.1E-04 | 9.6E-04 | \* |
| ENSG00000012817.16 | KDM5D | 5.9E+00 | 2.1E-03 | 7.1E-03 | \* |
| ENSG00000129824.16 | RPS4Y1 | 6.4E+00 | 4.6E-03 | 1.4E-02 | \* |
| ENSG00000198692.10 | EIF1AY | 6.4E+00 | 4.9E-03 | 1.5E-02 | \* |
| ENSG00000177257.3 | DEFB4B | 7.2E+00 | 2.1E-03 | 7.3E-03 | \* |

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| **Supplementary Table 2: Top 10 Significant Genes when comparing statistical models with all study participants (Model A) and study participants from Pueblo only (Model P).** | | | | | | | | |
| **Ensembl ID** | **Gene Symbol** | **Gene Type** | **Log2(FC) (Mod A)** | **Log2(FC) (Mod P)** | **Log2(FC) Difference (P - A)** | **Log2(FC) Difference (%)** | **FDR (Mod A)** | **FDR (Mod P)** |
| ENSG00000188039 | *NWD1* | protein coding | -2.2 | -2.2 | -0.1 | 2.5 | < 0.001 | < 0.001 |
| ENSG00000118997 | *DNAH7* | protein coding | -2.1 | -2.3 | -0.2 | 11.4 | < 0.001 | < 0.001 |
| ENSG00000105519 | *CAPS* | protein coding | -2.0 | -2.2 | -0.2 | 9.2 | < 0.001 | < 0.001 |
| ENSG00000287189 | *AL121956.6* | lncRNA | -2.8 | -3.1 | -0.3 | 9.8 | < 0.001 | < 0.001 |
| ENSG00000174844 | *DNAH12* | protein coding | -2.2 | -2.6 | -0.4 | 17.6 | < 0.001 | < 0.001 |
| ENSG00000154479 | *CCDC173* | protein coding | -2.4 | -2.6 | -0.2 | 9.0 | < 0.001 | < 0.001 |
| ENSG00000163263 | *C1orf189* | protein coding | -2.1 | -2.4 | -0.3 | 13.1 | < 0.001 | < 0.001 |
| ENSG00000197653 | *DNAH10* | protein coding | -2.4 | -2.7 | -0.3 | 12.3 | < 0.001 | < 0.001 |
| ENSG00000186976 | *EFCAB6* | protein coding | -2.2 | -2.5 | -0.2 | 10.1 | < 0.001 | < 0.001 |
| ENSG00000172955 | *ADH6* | protein coding | -2.9 | -3.3 | -0.4 | 14.5 | < 0.001 | < 0.001 |

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| **Supplementary Table 3: Top 10 Targeted CpG Sites by p-value based on the targeted epigenome wide analysis** | | | | | | | |
| CpG | Associated Gene Name | Estimate (M) | Std. Error | t value | p-value | FDR | Associated Gene Reg. |
| cg02123174 | REXO1 | −0.3 | 0.07 | −5.2 | < 0.001 | 0.01 | + |
| cg11903190 | CERK | 0.3 | 0.06 | 4.5 | < 0.001 | 0.06 | + |
| cg11171825 | PEX6 | −0.5 | 0.11 | −4.2 | < 0.001 | 0.16 | - |
| cg05241917 | ACE2 | −0.5 | 0.11 | −4.2 | < 0.001 | 0.16 | - |
| cg16232530 | C7orf50 | −0.4 | 0.10 | −4.3 | < 0.001 | 0.16 | - |
| cg06790566 | CDH22 | −0.3 | 0.08 | −4.4 | < 0.001 | 0.16 | - |
| cg04587192 | SRGAP3 | −0.3 | 0.08 | −4.3 | < 0.001 | 0.16 | - |
| cg23918706 | ZC2HC1C | 0.1 | 0.04 | 4.2 | < 0.001 | 0.16 | - |
| cg23918706 | ACYP1 | 0.1 | 0.04 | 4.2 | < 0.001 | 0.16 | - |
| cg15994418 | RAB35 | 0.2 | 0.05 | 3.9 | < 0.001 | 0.16 | + |

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| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Supplementary Table 4: Top differentially methylated regions (DMRs) by Sidak-adjusted p-values** | | | | | | | | |
| **Chromosome on which the DMR occurs** | **Starting base of the DMR** | **Ending base of the DMR** | **Sidak-adjusted p-value** | **# of CpG probes that occur within the DMR** | **List of CpG sites that occur within the DMR** | **Associated UCSC gene names** | **Number of positively methylated probes in DMR** | **Number of negatively methylated probes in DMR** |
| chr2 | 3,286,338 | 3,286,686 | <0.001 | 4 | cg13300473;cg15775218;cg15913725;cg25411977 | *EIPR1* | 4 | 0 |
| chr7 | 113,350,088 | 113,350,139 | <0.001 | 1 | cg14743902 | *PPP1R3A* | 0 | 1 |
| chr13 | 88,328,009 | 88,328,522 | <0.001 | 5 | cg05757365;cg09823859;cg16787483;cg24626752;cg26168643 | *SLITRK5* | 5 | 0 |
| chr1 | 19,971,709 | 19,972,201 | 0.004 | 5 | cg03884082;cg10211745;cg15589641;cg18923740;cg19234140 | *NBL1;MICOS10-NBL1* | 5 | 0 |
| chr12 | 132,639,764 | 132,640,226 | 0.005 | 3 | cg09451320;cg09957073;cg15321238 | *NOC4L* | 1 | 2 |
| chr8 | 101,225,245 | 101,225,953 | 0.006 | 6 | cg05800758;cg06790425;cg11067736;cg13687825;cg17103929;cg20270320 | *SPAG1* | 0 | 6 |
| chr2 | 240,270,992 | 240,271,327 | 0.007 | 5 | cg03585122;cg16299358;cg18803701;cg19770748;cg22009751 | *HDAC4* | 5 | 0 |
| chr18 | 14,748,040 | 14,748,349 | 0.014 | 7 | cg03014326;cg13266435;cg17014927;cg21281009;cg21293934;cg23703062;cg24061208 | *ANKRD30B* | 7 | 0 |
| chr6 | 74,364,495 | 74,364,751 | 0.018 | 4 | cg09416149;cg11742202;cg13599248;cg15842366 | *SLC17A5* | 4 | 0 |
| chr15 | 74,495,109 | 74,495,452 | 0.02 | 6 | cg00075967;cg11787522;cg14275207;cg15162876;cg18717554;cg26774156 | *STRA6* | 0 | 6 |
| chr8 | 51,095,593 | 51,095,883 | 0.031 | 2 | cg01355823;cg23046386 | *SNTG1* | 0 | 2 |
| chr8 | 23,711,711 | 23,712,025 | 0.032 | 4 | cg01798185;cg04252011;cg14599908;cg26321986 | *STC1* | 0 | 4 |
| chr15 | 69,351,555 | 69,351,865 | 0.034 | 2 | cg11155172;cg13323256 | *NOX5* | 0 | 2 |
| chr2 | 239,628,789 | 239,629,025 | 0.036 | 4 | cg05634047;cg06624310;cg17487719;cg25556432 | *LOC100287387* | 0 | 4 |
| chr8 | 142,482,939 | 142,483,073 | 0.037 | 2 | cg19593660;cg22226804 | *MROH5* | 0 | 2 |
| chr19 | 1,823,032 | 1,823,083 | 0.037 | 1 | cg02123174 | *LOC100288123;REXO1* | 0 | 1 |
| chr4 | 1,512,820 | 1,513,310 | 0.037 | 5 | cg03555012;cg04600233;cg11174255;cg18063486;cg25034424 | *NKX1-1* | 5 | 0 |
| chr19 | 46,056,709 | 46,056,958 | 0.045 | 4 | cg00767269;cg13588954;cg18489009;cg27391679 | *OPA3* | 4 | 0 |
| chr10 | 29,698,248 | 29,698,736 | 0.055 | 7 | cg03496709;cg04769618;cg14970695;cg16162391;cg17354476;cg19109608;cg26672999 | *SVIL-AS1* | 7 | 0 |
| chr15 | 89,960,503 | 89,960,794 | 0.056 | 3 | cg06059461;cg21760666;cg24871165 | *LOC105371031* | 3 | 0 |
| chr5 | 149,887,461 | 149,887,838 | 0.061 | 5 | cg06677890;cg07672051;cg12534263;cg14873515;cg16674484 | *NDST1* | 5 | 0 |
| chr16 | 1,797,204 | 1,797,434 | 0.076 | 2 | cg03650551;cg09080909 | *MAPK8IP3* | 0 | 2 |
| chr11 | 5,959,780 | 5,960,264 | 0.083 | 5 | cg03094675;cg05234568;cg13902645;cg14980045;cg25319279 | *OR56A3* | 5 | 0 |
| chr12 | 130,554,730 | 130,555,142 | 0.089 | 6 | cg04137484;cg06370094;cg07070348;cg09288218;cg17580782;cg25253445 | *LINC02419* | 0 | 6 |
| chr1 | 75,198,403 | 75,199,014 | 0.091 | 8 | cg02709834;cg07399417;cg09502221;cg10128416;cg13528603;cg21906852;cg26690034;cg26855724 | *TYW3;CRYZ* | 8 | 0 |
| chr20 | 60,294,656 | 60,294,991 | 0.091 | 4 | cg04614651;cg07123344;cg19921985;cg22031664 | *CDH4;LOC100128310* | 0 | 4 |
| chr19 | 1,151,960 | 1,152,409 | 0.094 | 5 |  | *SBNO2* | 5 | 0 |

Chart, box and whisker chart

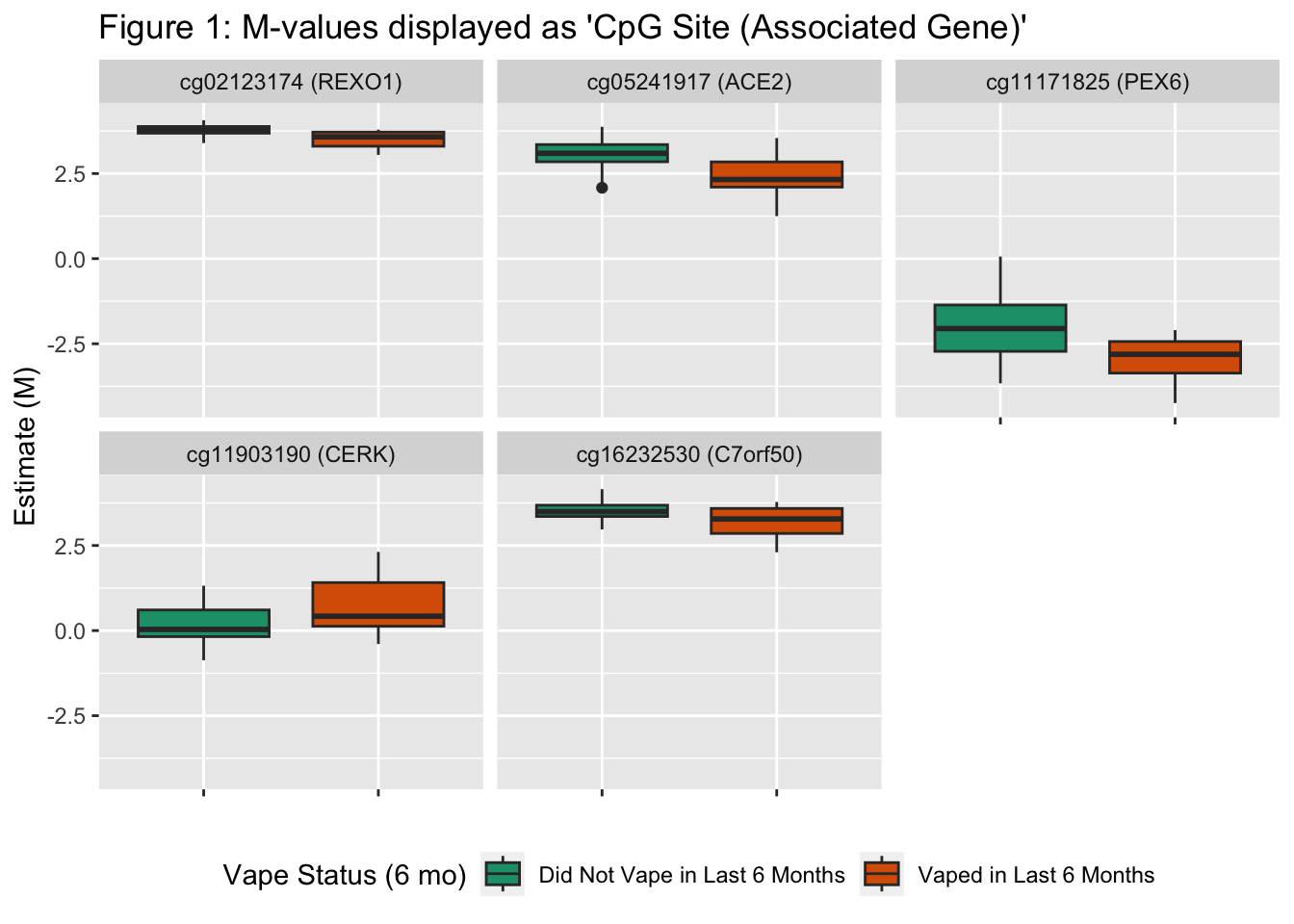
Description automatically generated

**Supplementary Figure 1. Differential expression of select genes between vape users and non-vape users. These genes were selected by filtering for only genes with |log2fc| > 2 then sorted by their p-values to define the ‘top’ up- and down-regulated genes.**

Chart, scatter chart

Description automatically generated

**Supplementary Figure 2. Significant genes from the model containing all study participants (*Model A)* and the model with only participants from the Pueblo recruitment center (*Model P)*.**



**Supplementary Figure 3. Boxplots of estimated relative methylation when vape users are compared to non-vape users.**